



## MicroRNA-626 inhibits mTOR pathways activity of retinal pigment epithelial cells by targeting SLC7A5 in human ARPE-19 Cells

Çilem Ercan<sup>1\*</sup>, Ahmet Elbay<sup>2</sup>, Elif Sibel Aslan<sup>3</sup>, Fahri Akbaş<sup>4</sup>, Hakan Ozdemir<sup>5</sup>, Nehir Ozdemir Ozgentürk<sup>6</sup>

<sup>1</sup> Faculty of Art and Science, Molecular Biology and Genetics, Yıldız Technical University, Istanbul, Turkey

<sup>2</sup> Bezmialem Vakıf University, Faculty of Medicine, Department of Ophthalmology, Istanbul, Turkey

<sup>3</sup> Department of Molecular Biology and Genetics, Biruni University, Topkapı, İstanbul, 34010, Turkey

<sup>4</sup> Bezmialem Vakıf University, Faculty of Medicine, Department of Medical Biology, Istanbul, Turkey

<sup>5</sup> Bezmialem University, Faculty of Medicine, Department of Ophthalmology, Istanbul, Turkey

<sup>6</sup> Faculty of Art and Science, Molecular Biology and Genetics, Yıldız Technical University, Istanbul, Turkey

### ARTICLE INFO

#### Original paper

#### Article history:

Received: March 12, 2023

Accepted: July 05, 2023

Published: October 31, 2023

#### Keywords:

AMD, ARPE-19, mir-626, mTOR, RPE, SLC7A5

### ABSTRACT

Recent studies have shown that miRNAs are associated with the pathological process involved in age-related macular degeneration (AMD). However, the microRNA-mediated post-transcriptional regulation in human retinal pigment epithelium (RPE) cells has not been adequately investigated. We investigated how miR-626 inhibits mTOR activity pathways and pathway-related genes in retinal pigment epithelial cells by targeting the solute carrier family seven-member 5 (SLC7A5) in ARPE19 cells. We transfected mir-626 mimic, mir-626 inhibitor and siRNA in human retinal pigment epithelial cell line was examined using RT-PCR and western blot, respectively. We knocked down mir-626 levels and overexpression by mir-626-siRNA transfection of human RPE cell lines, and using an MTT assay, we assessed the role of SLC7A5 on RPE cell proliferation. We additionally measured the expression of mTOR, Akt1, caspase 3, Bax, SLC17A7, SLC17A8, Creb1, Pten, HIF1A, HIF2A. The findings demonstrate that mir-626 inhibits SLC7A5 gene expression and proliferation of ARPE-19 cells. Short interfering RNA (siRNA) mediated suppression of SLC7A5, a predicted target of mir-626, has the same effect on ARPE-19 cells. We identified how miR-626 causes apoptosis and macula degeneration in RPE cells by targeting SLC7A5 through the mTOR signaling pathway. miR-626 was an essential regulator of the expression of the SLC7A5 gene. Importantly, we determined that miR-626 is essential to play a role in AMD. This research project shows that SLC7A5 is a direct target of mir-626 in ARPE-19 cells for the first time.

Doi: <http://dx.doi.org/10.14715/cmb/2023.69.10.3>

Copyright: © 2023 by the C.M.B. Association. All rights reserved.

### Introduction

Age-related macular degeneration is a multifactorial disease associated with a complex interaction of genetic and environmental factors. The early stage of AMD is characterized by dysfunction of RPE cells, with pigment changes and drusen deposition, which is essential for the function of the photoreceptors. In contrast, late-stage disease develops neovascularization and/or geographic atrophy with significant vision loss (1,2). Emerging evidence suggests that besides environmental and genetic factors, epigenetic mechanisms, such as microRNA (miRNA) regulation of gene expression, are relevant to AMD, providing an exciting new avenue for research and therapy (3). Some in vivo and in vitro studies suggested that numerous miRNAs are associated with the pathological process involved in AMD and diabetic retinopathy, such as pathological angiogenesis, oxidative stress, and inflammation, which indicates that these miRNAs might be potential therapeutic targets (4-6). The function and effects of miRNAs are reasonably well understood in the cell by recent studies and are promising for future miRNA-based studies. Furthermore, dysregulated miRNAs could be a treatment op-

tion by using miRNA mimics or antagomirs to modulate miRNA levels in the cell (7,8).

Our previous study showed that circulating miR-626 was significantly higher expression of serum miRNAs in patients with AMD (9). SLC7A5 is a molecular target of mir-626 demonstrated according to the MirTarbase.

SLC7A5 alias L type amino acid transporter (LAT1) (6) is a sodium-independent high-affinity amino acid transporter and mediates cellular uptake of the large neutral amino acids such as phenylalanine, tyrosine, leucine, and tryptophan (10). Leucine is an anabolic amino acid that stimulates the protein kinase, mammalian rapamycin (mTOR) target, protein translation, and cell growth (11-18). mTOR is the catalytic component of two complexes, the mammalian target of rapamycin complex 1 (mTORC1) and the mammalian target of rapamycin complex 2 (mTORC2). Also, Xu et al (19) have shown that SLC7A5 knockdown also decreased mTOR pathway activity (20).

The serine-threonine protein kinase Akt is a common mediator of cell survival signals. Akt signalling targets mTOR, which promotes angiogenesis. Failure of Akt-mediated signalling can cause apoptosis, leading to photoreceptor degeneration, bruch membrane thickening,

\* Corresponding author. Email: [cilem.ercan34@gmail.com](mailto:cilem.ercan34@gmail.com)

extracellular deposits, and decreased permeability leading to RPE damage, causing AMD.

Jomarv et al. demonstrated that in rd mice, inactivation of the Akt survival pathway results in photoreceptor cell death (21). Zhao et al. showed that mTOR-mediated dedifferentiation of the RPE indicates photoreceptor degeneration in mice (22). Therefore, we decided to examine mir 626 inhibiting mTOR pathway-related activity of retinal pigment epithelial cells by targeting SLC7A5 to understand the mTOR pathway-related activity molecular mechanism in the RPE of AMD. These findings were also confirmed by looking at mTOR, Akt1, caspase 3, Bax, SLC17A7, SLC17A8, Creb1, Pten, HIF1A, HIF2A, which are genes involved in pathways associated with macular degeneration.

## Materials and Methods

### Pathway Analyses

Target prediction and functional annotation were carried out as described in detail previously (9) that were performed using the miRSystem database (version 20160513) to evaluate the functions of candidate miRNAs of AMD patients, an integrated system for characterizing enriched processes.

### Cell Culture and Transfection

The human ARPE-19 cell line was purchased from ATCC (Manassas, VA). The cells were grown in Dulbecco's modified Eagle's medium and Ham's F12 medium (DMEM/F12) supplemented with 10% FBS (Hyclone, Logan, Utah), 100 U/ml of penicillin, and 100 µg/ml of streptomycin (Invitrogen, Gibco, Carlsbad, CA) at 37 °C under a humidified 5% CO<sub>2</sub>: 95% air atmosphere. The media has been changed once every three days. Synthesi-

zed RNA duplexes of miR-626 mimic, miR-626 inhibitor and short interfering RNAs (siRNAs) targeting SLC7A5 (AM1620) were purchased from Ambion (Austin, Texas). ARPE19 cells were seeded in 12-well plates at 1.5610 5 cells/well and cultured for 48 h and then transfected with scrambled miR-626 mimics, miR-626 inhibitor, short interfering RNAs (siRNAs) targeting SLC7A5 (AM1620) and controls at a final concentration of 50 nM in all experiments using Lipofectamine 2000 and OPTI-MEM I (Invitrogen Life, Technologies, Carlsbad, CA) and further incubated for 72 h before harvesting for RNA and protein analyses according to the manufacturer's protocol. Cells were incubated with the transfection complexes for 6h before replacing the medium. Cells were replaced with fresh growth medium daily (23).

### Total RNA Extraction and qRT-PCR

Total RNA was isolated using a Direct-zol™ RNA MiniPrep Kit (Zymo Research, Irvine, U.S.A) in accordance with the manufacturer's protocol. The qRT-PCR was performed following our previous descriptions (9). For quantification of mRNAs, mature miR-626 and RNU6, reverse transcription was performed using cDNA synthesis using the qScript cDNA Synthesis kit (Quanta Biosciences, Canada).

RT primers for mature miR-626 and RNU38B were supplied by SYBR Green reverse-transcription quantitative PCR performed with CFX Bio-Rad connect™ Real-Time Detection System (California, USA). Both groups' samples were analyzed in duplicate. The PCR program used for amplification was: 95 °C for 10 min, followed by 40 cycles of 95 °C for 15 s, 60 °C for 30 s, 72 °C for 45 s. GAPDH was used for normalization. PCR primer sequences are in Table 1. The results of mRNA and miRNAs were expressed as ΔCT, and fold changes in relative

**Table 1.** PCR Primer Sequences.

Gene Names	Primer Sequences
SLC7A5 Forward	5' ACGGCCGTGAAGTCTAC 3'
SLC7A5 Reverse	5' GGATCTAGATTGCGCAGAGGCCAGAGTT 3'
Akt1 Forward	5' CATCACACCACCTGACCAAT 3'
Akt1 Reverse	5' CTCTAAAATGCACCCGAGAAAAT 3'
Bax Forward	5' ATCCAGGATCGAGCAGGGCG 3'
Bax Reverse	5' GGTCTGATCAGTCCGGGCA 3'
Pten Forward	5' TGAGTTCCCTCAGCCGTTACCT 3'
Pten Reverse	5' GAGGTTTCTCTGGTCCTGGTA 3'
Caspase 3 Forward	5' TGCCTGTAAGTGTAGATGG 3'
Caspase 3 Reverse	5' CTCACCTTCTTACTTGGCGATGG 3'
HIF1A Forward	5' GAAACCACCTATGACCTGC 3'
HIF1A Reverse	5' CTGTTTGTGAAGGGAGAA 3'
HIF1A Forward	5' CCTGGCCATCAGCTTCCT 3'
HIF2A Reverse	5' GGTCGGCCTCAGCTTCAG 3'
Mtor Forward	5' ACCAGTGTGAGACCGTTTCC 3'
Mtor Reverse	5' AGGCAGGACTGGTGTATTGG 3'
Creb 1 Forward	5' GACCACTGATGGACAGCAGATC 3'
Creb 1 Reverse	5' GAGGATGCCATAACAACCTCCAGG 3'
Antisense SLC7A5	5' AACGGCGTGGCCATCATCGTGCCTGTCTC 3'
Sense SLC7A5	5' AACACGATGATGATGGCCACGCCGCTGTCTC 3'
GAPDH Forward	5' ACT CCA CTC ACG GCA AAT TC 3'
GAPDH Reverse	5' CAGTAGACTCCACGACATACT C 3'
hsa-mir-626	AGCUGUCUGAAAAUGUCUU
SLC17A7 Forward	5' GCAAGTACATCGAGGACGCCAT 3'
SLC17A7 Reverse	5' GCCACGATGATGGCATAGACTG 3'
SLC17A8 Forward	5' ACCACCTTTGGAGAGAAGCCGA 3'
SLC17A8 Reverse	5' GGACCATCCAATGTACTGCACC 3'

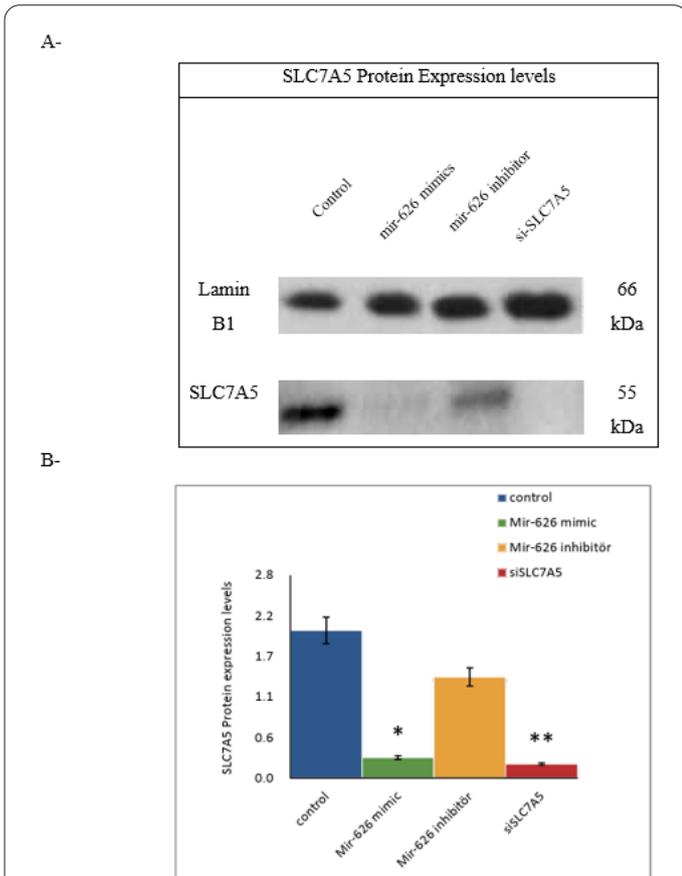


mimics (20 nM) from ARPE-19 cells as compared to mir-626 inhibitor (20 nM) from ARPE-19 cells ( $p < 0.005$ ) (Figure 2B). The mRNA expression levels of SLC7A5 were down-regulated in the siSLC7A5 group compared with the control group ( $p < 0.05$ ).

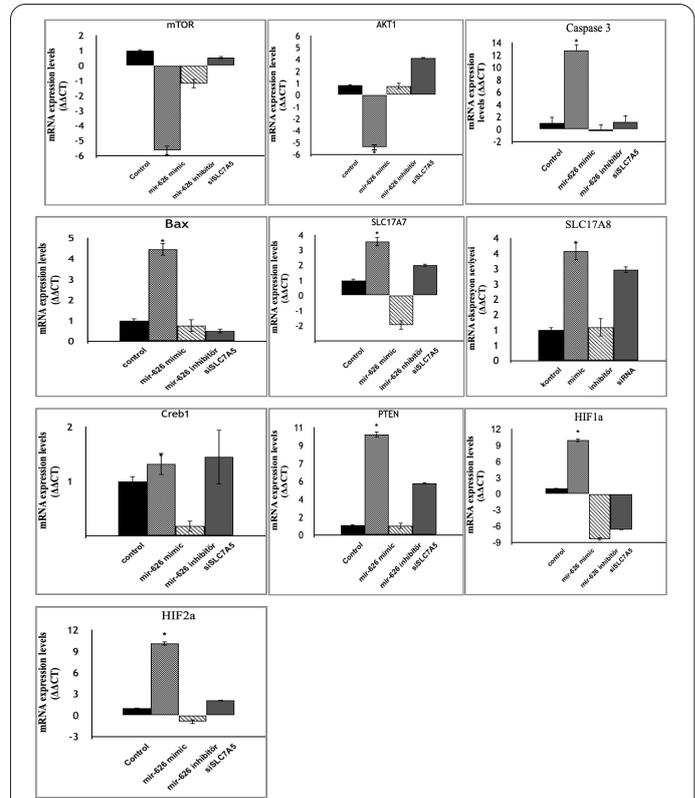
The western blot analyses further confirmed the qRT-PCR assay results. Our result showed that the protein expression levels decreased in the mir-626 mimics group compared with the control group ( $p < .05$ ), as you see in Figure 3A-3B. Transfection with the inhibitor of miR-626 resulted in no significant change in SLC7A5 protein expression ( $p > .05$ ) compared with the control, whereas the inhibitor of miR-626 protected the miR-626 targeting of SLC7A5. Also, the siSLC7A5 group was shown the same result as the mir-626 mimics group ( $p < .05$ ).

**Akt-mediated signalling mTOR pathway, angiogenesis and apoptosis pathways genes mRNA levels**

According to the results of Rt-PCR (Figure 4), with the control group, significant down-regulated was observed in the mRNA levels of mTOR, Akt, SLC17A7 in the mir-626 mimics group ( $p > .05$ ). The mir-626 mimics group, compared with the control group, significantly upregulated Bax, PTEN, Caspase-3, HIF1a, HIF1a and SLC17A8 mRNA levels ( $P < .05$ ). There were no significant differences in the mRNA levels of Creb1 in the mir-626 mimic



**Figure 3.** Effects of miR-626 mimics and inhibitors on SLC7A5 protein expression in ARPE-19 cells. **A-** Western Blot images of SLC7A5 protein levels in ARPE-19 cells of different groups: control, mir-626 mimics, mir-626 inhibitors, siSLC7A5. Values are presented as mean SEM; n = 3. \*p,0.05 vs. control. **B-** Quantification of relative SLC7A5 protein levels. The band intensity was measured by using ImageJ software (see ‘Materials and Methods’). The data reported here are representative of the experiment performed in triplicate. Values are presented as mean SEM; n = 3. \*p,0.05 vs. control.



**Figure 4.** miR-626 targets SLC7A5 to affect the Akt-mTOR pathway and Angiogenesis pathway in ARPE-19 cells. mTOR, Akt1, caspase 3, Bax, SLC17A7, SLC17A8, Creb1, Pten, HIF1A, and HIFI2A mRNA expression in ARPE-19 cells was detected by qRT-PCR. 1–4 represented control group, miR-626 mimics, miR-626 inhibitors group and siSLC7A5 group respectively. Each experiment was repeated three times. Data are presented as the mean ± SD. Multiple-group comparisons were analyzed by T-test. \*p < 0.05 compared with the control group and the miR-626 mimic group or compared with the miR-626 inhibitor group or compared with the siSLC7A5 group.

group compared with the control group ( $p > .05$ ).

**Discussion**

miRNAs are key regulators of several biological processes, such as pathological angiogenesis and the response to oxidative stress, and miRNA dysregulation has been linked with numerous diseases. miRNAs are involved in AMD pathology, and several miRNAs target genes and signaling pathways were identified concerning AMD pathogenesis and progression (24). Concerning AMD, recent miRNA studies focused on therapeutic research but have not shown any specific miRNAs directly linked with the pathology of AMD. For example, miR-126 can control vascular integrity and angiogenesis, which may provide a novel target for neovascular AMD (4). Our previous study showed that circulating mir-626, highly regulated and validated in sera from AMD patients, maybe a potential biomarker. Therefore, the bioinformatic database using mirTarbase showed that the human SLC7A5 3'UTR has three binding sites for the mir-626 (9). On the other hand, no studies were focusing on the mechanism of miR-626 angiogenesis and apoptosis in AMD by targeting specific genes.

To investigate the function of overexpressing the mir-626 in RPE cells, we focused on the SLC7A5 and hypothesized that the mir-626 target of SLC7A5 may modulate

via mTOR pathways activity to cause macula degeneration. Our data shows that mir-626 mimics and siRNA (siSLC7A5) suppressed the mRNA expression level and the protein level of SLC7A5 significantly.

In addition, recent studies have shown that the SLC7A5 gene is directly related to the mTOR pathway. Sokolov et al. demonstrated that Slc7a5 is required for mTOR pathway activity, maturation and survival, which may help explain why Slc7a5 mutations prevent normal brain development and function (25). Also, another recent report showed overexpressed SLC7A5 promotes mTOR-P70S6K signals and enhances the expression of MMP3 and MMP13 at the protein level in rheumatoid arthritis synoviocytes (20).

Akt signalling targets mTOR, which promotes angiogenesis (4). Jomarv et al. demonstrated that in mice, inactivation of the Akt survival pathway results in photoreceptor cell death (21). Our results showed that AKT1 mRNA expression levels were significantly decreased, and accordingly, mTOR mRNA expression levels were also significantly low expression in mir-626 mimics ARPE-19 cells compared to control cells.

The phosphatidylinositol 3-kinase (PI3K)/AKT pathway is directed downstream by the target of PTEN, PI3K signalling is suppressed, and pathway signalling does not occur. PI3K is involved in several cellular processes, including cell proliferation, apoptosis, and differentiation (26). Also, our data showed that Pten mRNA expression is significantly increased in mir-626 mimic groups and siSLC7A5 groups.

Elorza et al. showed that activation of the HIF2A pathway increases mTORC1 activity by upregulating the expression of the amino acid carrier SLC7A5 (27). Also, a recent study by Sokolov et al. reported that Slc7a5 is required for mTORC1 pathway activity, dendrite maturation, and survival (25). However, our result showed HIF1A and HIF2A mRNA expression levels were significantly upregulated in mir-626 mimics ARPE-19 cells compared to control cells. This study provides evidence that despite the increased expression of HIF1A and HIF2A genes, SLC7A5 gene expression and mTOR pathway activity in the cell ceased due to suppression of SLC7A5 by mir-626.

Another interesting fact is that many of these proteins are associated with the HIF-1 signaling pathway. HIF-1 is a transcriptional regulator that mediates the cellular responses to reduced oxygen levels through changes in gene expression (28).

Santos and colleagues came to the following conclusion in a study they conducted; glycolytic enzymes and glucose transporters appear to be upregulated in response to HIF-1 (29). HIF-1 $\alpha$  regulates SLC17A7 (Glut 1) and SLC17A8 (Glut 3). These data confirm our high expression levels of SLC17A7 and SLC17A8 mRNA.

The transcription factor CREB plays an important role in regulating cellular responses, like proliferation and survival, across multiple cell types exposed to oxidative stress (30). Creb anti-apoptotic signalling might inhibit caspase 3 activity and other cell death reactions (31). Our result demonstrated that Creb mRNA levels showed no significant change.

Some reports have revealed that H<sub>2</sub>O<sub>2</sub>-induced ARPE-19 cells apoptosis is related to the mitochondrial apoptotic signalling, which involves the proapoptotic protein Bax and the downstream protein caspase-3 (32-34). Bax and caspase3 mRNA expression levels were examined to un-

derstand the mir-626 role in the apoptosis pathway.

SLC7A5 knockdown by mir-626 significantly increases the Bax and caspase-3 mRNA expression levels in the apoptosis pathway in the mir-626 mimics group. In summary, our work has identified miR-626 as an essential regulator of the expression of the SLC7A5 gene. Thus, we identified how miR-626 causes apoptosis and macula degeneration in RPE cells by targeting SLC7A5 through the mTOR signaling pathway. Importantly, we determined that miR-626 is essential to play a role in AMD. However, more research is needed to understand the mechanism of miR-626 targeting SLC7A5 in RPE cells through the AKT- mTOR signaling pathway.

### Acknowledgement

The authors would like to acknowledge that this paper is submitted in partial fulfilment of the requirements for the Ph.D. degree at Yildiz Technical University.

### Funding

Founded by Bezmialem Vakıf University Scientific Research Project Office.

### Disclosure of potential interest conflicts

None of the authors has any potential conflict of interest.

### Contribution

Involved in the design and conduct of the study (CE, AE, NOO); data collection (CE, AE); analyse of the data (CE, FA, NOO); writing the article (CE, ESA, NOO); review of the study (CE, ESA, NOO, HA).

### References

1. Chen Y, Zeng J, Zhao C, et al. Assessing susceptibility to age-related macular degeneration with genetic markers and environmental factors. *Arch Ophthalmol* 2011;129:344–351.
2. Hogg RE, Chakravarthy U. Visual function and dysfunction in early and late age-related maculopathy. *Prog Retin Eye Res* 2006;25:249–276
3. Berber P, Grassmann F, Kiel C, Weber BH. An Eye on Age-Related Macular Degeneration: The Role of MicroRNAs in Disease Pathology. *Mol Diagn Ther*. 2017 Feb;21(1):31-43. doi: 10.1007/s40291-016-0234-z.
4. Wang S, Koster KM, He Y, Zhou Q. miRNAs as potential therapeutic targets for age-related macular degeneration. *Future Med Chem* 2012;4:277–28
5. Zhou Q, Gallagher R, Ufret-Vincenty R, Li X, Olson EN, Wang S. Regulation of angiogenesis and choroidal neovascularization by members of microRNA-23~27~24 clusters. *Proc Natl Acad Sci USA* 2011;108:8287–8292.
6. Urbich C, Kuehnbacher A, Dimmeler S. Role of microRNAs in vascular diseases, inflammation, and angiogenesis. *Cardiovasc Res* 2008;79:581–588.
7. Zhao H, Wang J, Gao L, Wang R, Liu X, Gao Z, et al. MiR-NA424 Protects against permanent focal cerebral ischemia injury in mice involving suppressing microglia activation. *Stroke* 2013;44:1706–13.
8. Bonauer A, Carmona G, Iwasaki M, Mione M, Koyanagi M, Fischer A, et al. MicroRNA-92a controls angiogenesis and functional recovery of ischemic tissues in mice. *Science* 2009;324:1710–3
9. Elbay A, Ercan C, Akbas F, Bulut H, Ozdemir H. Three new circulating microRNAs may be associated with wet age-related macular degeneration. *Scand J Clin Lab Invest* 2019;79:388–394.

10. Lim BC, Cho KY, Lim JS, Lee RS, Kim HS, Kim MK, Kim JH, Woo YJ, Kim JK, Kim DK, Kim HI, Lee KW, Lee MC. Increased expression of L-amino acid transporters in balloon cells of tuberous sclerosis. *Childs Nerv Syst*. 2011 Jan;27(1):63-70. doi: 10.1007/s00381-010-1239-2.
11. Chen R, Zou Y, Mao D, Sun D, Gao G, Shi J, Liu X, Zhu C, Yang M, Ye W, Hao Q, Li R, Yu L. The general amino acid control pathway regulates mTOR and autophagy during serum/glutamine starvation. *J Cell Biol*. 2014 Jul 21;206(2):173-82. doi: 10.1083/jcb.201403009. PMID: 25049270; PMCID: PMC4107793.
12. Kimball SR, Gordon BS, Moyer JE, Dennis MD, Jefferson LS. Leucine induced dephosphorylation of Sestrin2 promotes mTORC1 activation. *Cell Signal*. 2016 Aug;28(8):896-906. doi: 10.1016/j.cellsig.2016.03.008. Epub 2016 Mar 21. PMID: 27010498; PMCID: PMC4899281.
13. Wolfson RL, Chantranupong L, Saxton RA, Shen K, Scaria SM, Cantor JR, Sabatini DM. Sestrin2 is a leucine sensor for the mTORC1 pathway. *Science*. 2016 Jan 1;351(6268):43-8. doi: 10.1126/science.aab2674.
14. Parmigiani A, Nourbakhsh A, Ding B, Wang W, Kim YC, Akopiants K, Guan KL, Karin M, Budanov AV. Sestrins inhibit mTORC1 kinase activation through the GATOR complex. *Cell Rep*. 2014 Nov 20;9(4):1281-91. doi: 10.1016/j.celrep.2014.10.019.
15. Sancak Y, Bar-Peled L, Zoncu R, Markhard AL, Nada S, Sabatini DM. Ragulator-Rag complex targets mTORC1 to the lysosomal surface and is necessary for its activation by amino acids. *Cell*. 2010 Apr 16;141(2):290-303. doi: 10.1016/j.cell.2010.02.024.
16. Saxton RA, Knockenhauer KE, Wolfson RL, Chantranupong L, Pacold ME, Wang T, Schwartz TU, Sabatini DM. Structural basis for leucine sensing by the Sestrin2-mTORC1 pathway. *Science*. 2016 Jan 1;351(6268):53-8. doi: 10.1126/science.aad2087.
17. Sancak Y, Peterson TR, Shaul YD, Lindquist RA, Thoreen CC, Bar-Peled L, Sabatini DM. The Rag GTPases bind raptor and mediate amino acid signaling to mTORC1. *Science*. 2008 Jun 13;320(5882):1496-501. doi: 10.1126/science.1157535. Epub 2008 May 22. PMID: 18497260; PMCID: PMC2475333.
18. Hara K, Yonezawa K, Weng QP, Kozlowski MT, Belham C, Avruch J. Amino acid sufficiency and mTOR regulate p70 S6 kinase and eIF-4E BP1 through a common effector mechanism. *J Biol Chem*. 1998 Jun 5;273(23):14484-94. doi: 10.1074/jbc.273.23.14484. Erratum in: *J Biol Chem* 1998 Aug 21;273(34):22160. PMID: 9603962.
19. Laplante M, Sabatini DM. mTOR Signaling. *Cold Spring Harb Perspect Biol*. 2012 Feb 1;4(2):a011593. doi: 10.1101/cshperspect.a011593. PMID: 22129599; PMCID: PMC3281571.
20. Xu J, Jiang C, Cai Y, Guo Y, Wang X, Zhang J, Xu J, Xu K, Zhu W, Wang S, Zhang F, Geng M, Han Y, Ning Q, Xu P, Meng L, Lu S. Intervening upregulated SLC7A5 could mitigate inflammatory mediator by mTOR-P70S6K signal in rheumatoid arthritis synovial cells. *Arthritis Res Ther*. 2020 Aug 31;22(1):200. doi: 10.1186/s13075-020-02296-8. PMID: 32867828; PMCID: PMC7457370.
21. Jomary C, Cullen J, Jones SE. Inactivation of the Akt survival pathway during photoreceptor apoptosis in the retinal degeneration mouse. *Invest Ophthalmol Vis Sci*. 2006 Apr;47(4):1620-9. doi: 10.1167/iovs.05-1176. PMID: 16565401.
22. Zhao C, Yasumura D, Li X. mTOR-mediated dedifferentiation of the retinal pigment epithelium initiates photoreceptor degeneration in mice. *J Clin Invest* 2011; 121(1), 369–383.
23. Miko E, Margitai Z, Czimmerer Z, Várkonyi I, Dezso B, Lányi A, Bacsó Z, Scholtz B. miR-126 inhibits proliferation of small cell lung cancer cells by targeting SLC7A5. *FEBS Lett*. 2011 Apr 20;585(8):1191-6. doi: 10.1016/j.febslet.2011.03.039. Epub 2011 Mar 23. PMID: 21439283.
24. Elshelmani, Hanan, and Sweta Rani. "Exosomal microRNA discovery in age-related macular degeneration." *MicroRNA Profiling*. Humana Press, New York, NY, 2017. 93-113.
25. Sokolov AM, Holmberg JC, Feliciano DM. The amino acid transporter Slc7a5 regulates the mTOR pathway and is required for granule cell development. *Hum Mol Genet*. 2020 Nov 4;29(18):3003-3013. doi: 10.1093/hmg/ddaa186. PMID: 32821949; PMCID: PMC7645712.
26. Kang BW, Chau I. Molecular target: pan-AKT in gastric cancer. *ESMO Open*. 2020 Sep;5(5):e000728. doi: 10.1136/esmoopen-2020-000728. PMID: 32948630; PMCID: PMC7511610.
27. Elorza A, Soro-Arnáiz I, Meléndez-Rodríguez F, Rodríguez-Vaello V, Marsboom G, de Cárcer G, Acosta-Iborra B, Albacete-Albacete L, Ordóñez A, Serrano-Oviedo L, Giménez-Bachs JM, Vara-Vega A, Salinas A, Sánchez-Prieto R, Martín del Río R, Sánchez-Madrid F, Malumbres M, Landázuri MO, Aragonés J. HIF2 $\alpha$  acts as an mTORC1 activator through the amino acid carrier SLC7A5. *Mol Cell*. 2012 Dec 14;48(5):681-91. doi: 10.1016/j.molcel.2012.09.017. Epub 2012 Oct 25. PMID: 23103253.
28. Vadlapatla RK, Vadlapudi AD, Mitra AK. Hypoxia-inducible factor-1 (HIF-1): a potential target for intervention in ocular neovascular diseases. *Curr Drug Targets*. 2013 Jul;14(8):919-35. doi: 10.2174/13894501113149990015. PMID: 23701276; PMCID: PMC4407697.
29. Santos FM, Gaspar LM, Ciordia S, Rocha AS, Castro E Sousa JP, Paradela A, Passarinha LA, Tomaz CT. iTRAQ Quantitative Proteomic Analysis of Vitreous from Patients with Retinal Detachment. *Int J Mol Sci*. 2018 Apr 11;19(4):1157. doi: 10.3390/ijms19041157. PMID: 29641463; PMCID: PMC5979392.
30. Sugiura S, et al. CRE-mediated gene transcription in the peri-infarct area after focal cerebral ischemia in mice. *J Neurosci Res* 2004;75(3):401–7
31. Fang J, Zhao X, Li S, Xing X, Wang H, Lazarovici P, Zheng W. Protective mechanism of artemisinin on rat bone marrow-derived mesenchymal stem cells against apoptosis induced by hydrogen peroxide via activation of c-Raf-Erk1/2-p90rsk-CREB pathway. *Stem Cell Res Ther*. 2019 Oct 26;10(1):312. doi: 10.1186/s13287-019-1419-2. PMID: 31655619; PMCID: PMC6815409.
32. Azizi Dargahlou, S., Iriti, M., Pouresmaeil, M., Goh, L. P. W. MicroRNAs; their therapeutic and biomarker properties. *Cell Mol Biomed Rep* 2023; 3(2): 73-88. doi: 10.55705/cnbr.2022.365396.1085
33. Kanwal, N., Al Samarrai, O., Al-Zaidi, H. M. H., Mirzaei, A., Heidari, M. Comprehensive analysis of microRNA (miRNA) in cancer cells. *Cell Mol Biomed Rep* 2023; 3(2): 89-97. doi: 10.55705/cnbr.2022.364591.1070.
34. Yan Y, Ren Y, Li X, Zhang X, Guo H, Han Y, Hu J. A polysaccharide from green tea (*Camellia sinensis* L.) protects human retinal endothelial cells against hydrogen peroxide-induced oxidative injury and apoptosis. *Int J Biol Macromol*. 2018 Aug;115:600-607. doi: 10.1016/j.ijbiomac.2018.04.011. Epub 2018 Apr 5. PMID: 29627466.