

Original Article

Salivary interleukin-17A and interleukin-18 as potential biomarkers of immunopathogenesis and oral health status in rheumatoid arthritis

Ahmed Abdul-hassan Abbas¹ , Zahraa F. Shaker², Batool Hassan Al-Ghurabi^{3*} , Ghaib Nidhal H⁴, Mohammed Ahmed Abdulhassan⁵, Mustafa Ahmed Abdulhassan⁴

¹ Department of Microbiology, College of Medicine, Al-Nahrain University, Baghdad, Iraq

² Department of Basic Science, College of Dentistry, University of Al-Mustansiriyah, Baghdad, Iraq

³ Department of Basic Science, College of Dentistry, University of Baghdad, Baghdad, Iraq

⁴ Department of Dentistry, Al-Esraa University, Baghdad, Iraq

⁵ Alkindy College of Medicine, University of Baghdad, Baghdad, Iraq

Article Info

Abstract



Article history:

Received: March 28, 2025

Accepted: July 08, 2025

Published: August 31, 2025

Use your device to scan and read the article online



Rheumatoid arthritis (RA) is a chronic autoimmune disease characterized by persistent inflammation and is often associated with poor oral health. Cytokines play a central role in RA immunopathogenesis. This case-control study investigated the involvement of salivary interleukin-17A (IL-17A) and interleukin-18 (IL-18) in RA patients in relation to oral health status. Unstimulated whole saliva samples were collected from 20 RA patients and 20 age- and sex-matched healthy controls. Oral health was assessed using plaque and gingival indices. Salivary IL-17A and IL-18 concentrations were measured by ELISA. RA patients exhibited significantly higher salivary levels of IL-17A and IL-18 compared to controls ($p < 0.05$). Both cytokines showed positive correlations with gingival index, and IL-17A was also correlated with disease activity (DAS28). Receiver operating characteristic (ROC) analysis demonstrated that both interleukins effectively discriminated RA patients from healthy controls (AUC = 0.927 for IL-17A, AUC = 0.925 for IL-18). These findings suggest that elevated salivary IL-17A and IL-18 are associated with increased oral inflammation and may serve as non-invasive biomarkers for RA immunopathogenesis and oral health deterioration.

Keywords: Rheumatoid arthritis, Oral health, Cytokines, IL-17A, IL-18.

1. Introduction

Rheumatoid arthritis (RA) is a systemic autoimmune disease characterized by chronic inflammation of the joints and various extra-articular tissues. Its development is influenced by a combination of genetic predisposition and environmental factors [1]. The initial stages of RA are marked by fatigue, signs resembling the flu, inflamed and sore joints, and stiffness in the morning, as well as raised levels of C-reactive protein and erythrocyte sedimentation rate [2]. Patients with arthritis often show oral signs, including a strong association with Sjögren's syndrome, which causes signs such as xerostomia. Other oral concerns include temporomandibular joint difficulties, methotrexate-induced ulcers, and an increased focus on gum disease [3]. There is presently a wealth of information describing and documenting the connection between dental health and overall health [4, 5, 6, 7]. Bad oral hygiene and/or low oral and periodontal health were prevalent in RA patients, and metrics like bleeds, gum inflammation, and the depth of pockets in the gums were frequently significant when

compared to normal subjects. Improvements in oral hygiene and early nonsurgical periodontal therapy can also lessen the severity of systemic diseases, according to clinical trials [8, 9]. Immune and non-immune cells communicated with one another through cytokines. Therefore, the pathophysiology of a number of diseases, including RA, depends on these cytokines [10, 11].

In 2003, the Th17 subgroup of CD4 T cells was shown to produce IL-17A. It has been linked to the etiology of a number of inflammatory and autoimmune conditions, including psoriasis, gum disease, RA, and inflammatory bowel disorders [12]. One pleiotropic cytokine that is crucial to the onset and maintenance of the inflammatory response in RA is IL-18. In order to mediate bone degradation, IL-18 activates T cells in the synovium to generate inflammatory cytokines, such as RANKL [13]. Salivary diagnostics have shown enormous promise in clinical applications, and saliva has been shown to be a promising body fluid for early illness detection [14]. Thanks to combinations of biomolecules with clinical relevance and im-

* Corresponding author.

E-mail address: batoolamms@codental.uobaghdad.edu.iq (B.I Hassan Al-Ghurabi).

Doi: <http://dx.doi.org/10.14715/cmb/2025.71.8.15>

provements in detection technology, saliva may soon be the preferred first-line diagnostic sample [15]. This study was conducted to ascertain the role of salivary cytokines (IL-17A and IL-18) in the immunopathogenesis of RA with regard to oral health status because it has recently been proposed that these levels play a significant role in the pathogenesis of both systemic and oral diseases.

2. Materials and Methods

2.1. Study design

This study was designed as a case-control investigation.

2.2. Study population

A total of 40 participants were enrolled, including 20 patients diagnosed with rheumatoid arthritis (RA) and 20 age- and sex-matched healthy controls. RA diagnosis and assessment of disease activity (DAS28) were performed by a specialist rheumatologist according to the 2010 ACR/EULAR criteria. Exclusion criteria included the presence of other autoimmune or systemic inflammatory diseases, smoking, alcohol consumption, pregnancy, use of medications affecting immune function, and lack of informed consent. Ethical approval was obtained from the Scientific Committee of the College of Dentistry, University of Baghdad.

2.3. Oral examination

Clinical oral assessments were conducted, including evaluation of the plaque index (PLI) and gingival index (GI), following established protocols [16].

2.4. Saliva collection

Unstimulated whole saliva was collected from each participant by passive drooling into a sterile disposable container, avoiding stimulation or spitting. Approximately 3 mL of saliva was obtained per subject. Samples were transferred to sterile tubes, centrifuged at $2,000 \times g$ for 3 minutes at 4°C , and the clear supernatant was aliquoted into Eppendorf tubes and stored at -20°C until analysis [17].

2.5. Measurement of interleukin levels

Salivary concentrations of IL-17A and IL-18 were measured using a sandwich enzyme-linked immunosorbent as-

say (ELISA) according to the manufacturer's instructions (Shanghai, China). Briefly, capture antibodies specific for each biomarker were immobilized on the wells of a strip plate. Samples and standards were added and incubated, followed by the addition of biotinylated secondary antibodies. After washing, HRP-conjugate was added, and color development was achieved using a chromogenic substrate. The reaction was stopped with sulfuric acid, and absorbance was measured at 450 nm using an ELISA plate reader. Cytokine concentrations were calculated based on standard curves.

2.6. Sample size calculation

Sample size was determined using G*Power 3.1.9.7 (Franz Faul, Universität Kiel, Germany) with a power of 95% and a two-sided alpha error of 0.05.

2.7. Statistical analysis

Statistical analyses were performed using SPSS version 25. Descriptive statistics were expressed as mean \pm standard deviation. The Shapiro-Wilk test was used to assess the normality of data distribution. The Chi-square test was applied for categorical variables. Comparisons between two groups were performed using the independent t-test or Mann-Whitney U test, as appropriate. Pearson and Spearman correlation coefficients were used to assess associations between clinical and cytokine parameters. Receiver operating characteristic (ROC) curve analysis was conducted to evaluate the diagnostic performance of salivary cytokines. A p-value < 0.05 was considered statistically significant.

3. Results

3.1. Demographic and clinical characteristics of study participants

Table 1 summarizes the demographic and clinical characteristics of the 40 participants in this study. The mean age of RA patients was 47.4 ± 7.63 years, compared to 46.05 ± 6.31 years in the control group. The female-to-male ratio was 13:7 in the RA group and 12:8 in the control group. There were no significant differences between the groups in terms of age or sex distribution ($P > 0.05$). The mean DAS28 score in the RA group was 4.1 ± 0.84 , indicating moderate disease activity. Additionally, both the plaque index (PLI) and the gingival index (GI) were signi-

Table 1. Demographic features and oral variables in two groups.

Demographic features	Study group	healthy group	P-value
Age			
Range	29-59	32-59	0.272 ^{NS#}
Mean \pm SD	47.4 \pm 7.63	46.05 \pm 6.31	
Sex			
Female	13 (65%)	12 (60%)	0.743 ^{NS##}
Male	7 (35%)	8 (40%)	
Oral health status			
PLI			
Mean \pm SD	1.61 \pm 0.49	0.43 \pm 0.05	<0.000* [#]
GI			
Mean \pm SD	1.70 \pm 0.65	0.95 \pm 0.22	<0.000* [#]
DAS28			
Mean \pm SD	4.1 \pm 0.84	-	-

^{NS}: non-significant; *: significant; SD: standard deviation; #: t-test; ##: chi-square.

ificantly higher in RA patients than in controls ($P < 0.05$), reflecting poorer oral health status in the patient group.

3.2. Salivary cytokine levels and their correlation with clinical parameters

Regarding salivary interleukin levels, our findings demonstrated that both the mean IL-17A and the median IL-18 concentrations were significantly higher in the RA group compared to the control group ($P < 0.05$; Table 2). Furthermore, in RA patients, the mean IL-17A level

showed significant positive correlations with both the gingival index (GI) and disease activity score (DAS28). Similarly, the median IL-18 level was significantly correlated with GI and with IL-17A levels (Table 3, Figure 1).

3.3. Diagnostic performance of salivary cytokines

To evaluate the sensitivity and specificity of salivary interleukins, a receiver operating characteristic (ROC) analysis was performed to distinguish RA patients from healthy controls. Both IL-17A and IL-18 demonstrated excellent discriminatory power, as shown in Figure 2 and Table 4. The area under the curve (AUC) values indicated

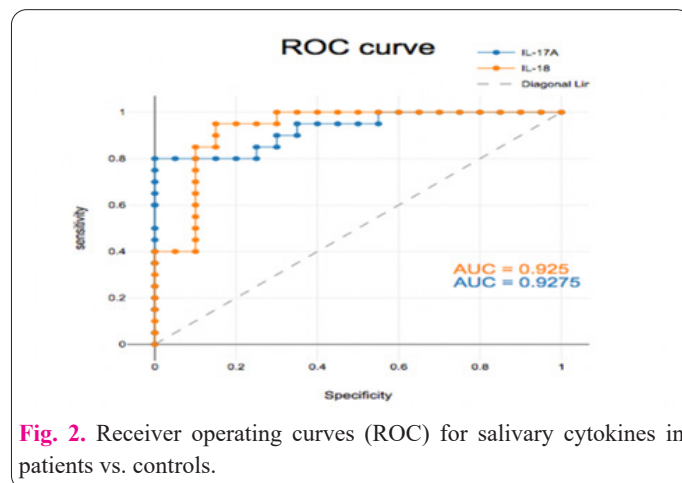
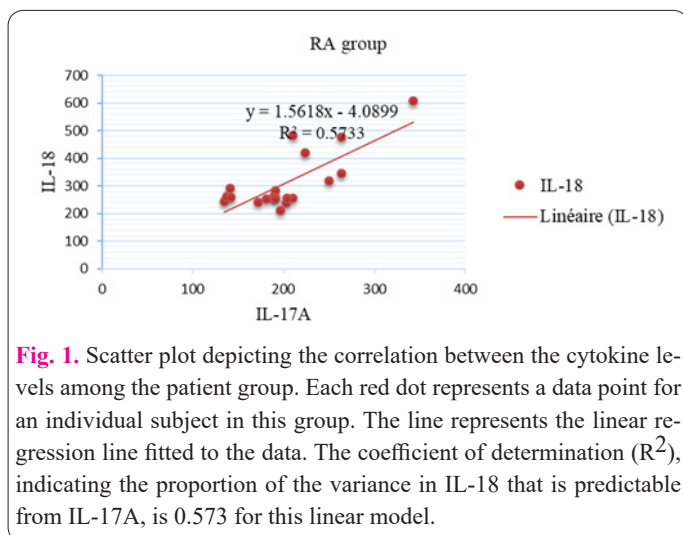


Table 2. Salivary concentrations of cytokines in the two groups.

Salivary cytokines	study group	healthy group	P-value
IL-17			
Min-Max	135-343	85.17-149.84	
Mean± SD	201.95±50.42	129±19.06	0.000*#
Median	194	135	
IL-18			
Min-Max	212.84-607.99	143.50-282.77	
Mean± SD	311.31±104.01	199.41±37.54	
Median	261.66	201.06	0.000*+

*: significant; SD: standard deviation; #: t-test; +: Mann-Whitney Test.

Table 3. Correlation between cytokines and clinical parameters in the patient group.

Pearson's correlation		RA group			
		IL-17A	PLI	GI	DAS28
IL-17A			r=0.228 p=0.332	r=0.485 p=0.030*	r=0.458 p=0.041*
PLI	r=0.228 p=0.332			r=0.283 p=0.225	r=0.088 p=0.711
GI	r=0.485 p=0.030*		r=0.283 p=0.225		r=0.227 p=0.335
DAS28	r=0.458 p=0.041*		r=0.088 p=0.711	r=0.227 p=0.335	
Spemann's correlation	RA group				
	PLI		GI	DAS28	IL-17A
IL-18	r=0.332 p=0.151	r=0.790 p=0.000*	r=0.368 p=0.110	r=0.547 p=0.012*	

Table 4. Comparison of the diagnostic properties of interleukins between the two groups.

Comparison	Test result variables (s)	AUC	P-value	Optimal cut-off point	sensitivity	specificity
RA Vs. Controls	IL-17A	0.927	0.000*	238	100%	%95
	IL-18	0.925	0.000*	172	100%	%95

that both interleukins are highly effective in differentiating between the two groups.

4. Discussion

The mean PLI and GI values in RA patients were increased as compared to healthy controls, according to this study. This indicates that the gingival tissue among patients was more inflamed, and it may be connected to the rise in plaque, which is the cause of gingival inflammation [18], which concurs with this outcome. GI and PLI are likely caused by increased likelihood of temporomandibular joint involvement in RA patients, significant arthritis-related hand malfunction that restricts the patient's ability to move and, concurrently, decreased saliva from secondary Sjogren's syndrome, both of which lead to plaque buildup. As well as RA patients' emotional depression about their condition, which deteriorates their attention to personal hygiene [18,19]. Saliva is now a widely accessible, non-invasive biofluid for identifying biomarkers in a number of conditions, including RA. A few advantages of using saliva for diagnostic purposes include ease of collection, reduced patient discomfort, and frequent monitoring. In the context of RA, salivary cytokines may provide valuable insights into the progression of illness and the efficacy of treatment [20].

The current results were consistent with a prior study [21, 22], which revealed that the RA patients had higher mean levels of IL-17A than the control group, and also observed that cytokine expression levels were out of balance, with RA patients having higher levels of IL-17 than controls. In addition, Atwa et al. [23] reported that when RA patients were compared to healthy controls, their levels of IL-17 were noticeably higher in RA cases. Moreover, Moran et al. [24] found that RA disease activity is driven by high production of IL-17A level in the inflamed joint. This supports our finding regarding the positive correlation between IL-17A and DAS28. Thus, evidence points to the significance of IL-17A during RA, emphasizing its potential for prognosis and disease surveillance. It is worth mentioning that studies conducted by Rosu et al. [25] and Jain and associates [26] showed that IL-17A is essential to the pathophysiology of RA. This could be the result of the inflammatory response. The immune system is overactive in those patients, which results in higher levels of IL-17A as part of the body's reaction to inflammation. Its elevation suggests the presence of an ongoing inflammatory process, which is particularly prominent in patients whose disease activity is high.

Similar to the results from numerous studies, the present results found that RA patients' group had statistically significant increase in median IL-18 level when compared to control group [27, 28], revealing that RA patients' IL-18 levels were higher than controls. Moreover, Shao et al. [29] showed that IL-18 biological activity in their blood, synovial fluid, and tissue than control groups. Broz and Dixit [30] reported that the activation of inflammasomes may be the cause of elevated salivary IL-18. To present various data, recent findings discovered that the median salivary IL-18 level showed no discernible difference between RA patients and controls [31]. This disparity could result from the distinct population under investigation or the various sample kinds.

The positive correlation between IL-17A and IL-18 in this study may indicate the involvement of these interleu-

kins in the inflammatory process in the periodontal tissues in RA patients. On the other hand, the positive correlation between interleukins (IL-17A and IL-18) and GI in RA patients, this could be due to the fact that increased production of inflammatory interleukins involved in the inflammation periodontal tissues in oral cavity and indicates that a large number of RA patients struggle with dental self-care and dental hygiene. Additionally, Salivary IL-17A and IL-18 demonstrated excellent clinical accuracy in discriminating RA patients from healthy individuals.

This study revealed significantly higher levels of salivary IL-17A and IL-18 in rheumatoid arthritis (RA) patients compared to healthy controls, with these elevations correlating positively with increased Plaque Index (PLI) and Gingival Index (GI) scores. Such associations underscore the interplay between systemic inflammation in RA and oral health status. These findings reinforce the established link between periodontal disease and RA, suggesting that poor oral health may exacerbate RA progression. Consequently, incorporating diligent oral hygiene practices and periodontal treatments into the standard care for RA patients is imperative to potentially mitigate both oral and systemic inflammatory burdens.

Conflicts of interest

The authors declare that they have no conflicts of interest.

Funding

No funding was received.

Authors' contributions

BHAG and AAA were involved in the conception and design of the study, in the literature search, clinical analysis, data analysis, statistical analysis, and in manuscript preparation and manuscript reviewing. All authors were involved in the conception and design of the study, in data analysis, and in manuscript preparation and manuscript reviewing. All authors have read and approved the final manuscript.

References

1. Klareskog L, Padyukov L, Lorentzen J, Alfredsson L (2006) Mechanisms of disease: Genetic susceptibility and environmental triggers in the development of rheumatoid arthritis. *Nat Clin Pract Rheumatol* 2(8):425–433 doi: <https://doi.org/10.1038/ncprheum0249>
2. Brzustewicz E, Henc I, Daca A, Szarecka M, Sochocka-Bykowska M, Witkowski J, Bryl E (2017) Autoantibodies, C-reactive protein, erythrocyte sedimentation rate and serum cytokine profiling in monitoring of early treatment. *Cent Eur J Immunol* 42:259–268 doi:10.5114/ceji.2017.70968
3. Benli M, Batool F, Stutz C, Petit C, Jung S, Huck O (2021) Orofacial manifestations and dental management of systemic lupus erythematosus: A review. *Oral Dis* 27(2):151–167 doi: <https://doi.org/10.1111/odi.13271>
4. Li R, Tian C, Postlethwaite A, et al. (2017) Rheumatoid arthritis and periodontal disease: what are the similarities and differences. *Int J Rheum Dis* 20(12):1887–1901 doi: <https://doi.org/10.1111/1756-185x.13240>
5. Daily ZA, Al-Ghurabi BH, Al-Qarakhli AM, Moseley R (2023) MicroRNA-155 (miR-155) as an accurate biomarker of periodontal status and coronary heart disease severity: a case-control study. *BMC Oral Health* 23(1):868 doi:10.1186/s12903-023-03584-w

6. Daily ZA, Al-Ghurabi BH, Al-Qarakhli AM, Hussein HM (2023) Association between AIM2 and pycard genes polymorphisms and susceptibility to periodontitis with coronary heart disease. *J Clin Cosmet Investig Dent* 15:307–320 doi:10.2147/CCIDE.S440577
7. Al Obaidi MJ, Al Ghurabi BH (2023) Potential role of NLRP3 inflammasome activation in the pathogenesis of periodontitis patients with type 2 diabetes mellitus. *J Med Chem Sci* 6:522–531 doi: <https://doi.org/10.26655/JMCHEMSCI.2023.3.9>
8. Fuggle NR, Smith TO, Kaul A, Sofat N (2016) Hand to mouth: a systematic review and meta-analysis of the association between rheumatoid arthritis and periodontitis. *Front Immunol* 7:80 doi: <https://doi.org/10.3389/fimmu.2016.00080>
9. Nori SA, Al Ghurabei BH (2025) Potential role of periodontopathogens in rheumatoid arthritis. *Dent* 3000 13(1):808 doi: <https://doi.org/10.5195/d3000.2025.808>
10. Al-Ghurabi BH (2021) The role of soluble TLR-2 in the immunopathogenesis of gingivitis. *Int Med J* 28:37–39
11. Aldaher Z, Al-Ghurabi B, Alwan B (2018) Serum levels of IL-22 and ACPA in patients with rheumatoid arthritis. *J Pure Appl Microbiol* 12:687–691 doi: <http://dx.doi.org/10.22207/JPaM.12.2.27>
12. Silva N, Dutzan N, Hernandez M, Dezerega A, Rivera O, Aguilon JC, et al. (2008) Characterization of progressive periodontal lesions in chronic periodontitis patients: levels of chemokines, cytokines, matrix metalloproteinase-13, periodontal pathogens and inflammatory cells. *J Clin Periodontol* 35:206 doi: <https://doi.org/10.1177/0022034511401405>
13. Fahey E, Doyle SL (2019) IL-1 family cytokine regulation of vascular permeability and angiogenesis. *Front Immunol* 10:1426 doi: <https://doi.org/10.3389/fimmu.2019.01426>
14. Lee YH, Wong DT (2009) Saliva: an emerging biofluid for early detection of diseases. *Am J Dent* 22(4):241 doi: <https://doi.org/10.1016/j.cden.2010.08.004>
15. Pfaffe T, Cooper-White J, Beyerlein P, Kostner K, Punyadeera C (2011) Diagnostic potential of saliva: current state and future applications. *Clin Chem* 57(5):675–687 doi: 10.1373/clinchem.2010.153767
16. Silness J, Loe H (1964) Periodontal disease in pregnancy. Correlate between oral hygiene and periodontal condition. *Acta Odontol Scand* 22:35 doi: 10.3109/00016356408993968
17. Navazesh M (1993) Methods for collecting saliva. *Ann N Y Acad Sci* 694(1):72–77 doi: <https://doi.org/10.1111/j.1749-6632.1993.tb18343.x>
18. Kässer UR, Gleissner C, Dehne F, Michel A, Boltz WW (1997) Risk for periodontal disease in patients with longstanding rheumatoid arthritis. *Arthritis Rheum* 40(12):2248–2251 doi: 10.1002/art.1780401221
19. Jarallah FM, Al-Safi K, Al-Ghurabi BH (2012) Evaluation of serum anti-cyclic citrullinated peptide antibodies level in rheumatoid patients with and without periodontitis. *J Bagh Coll Dent* 24:83–87
20. Kaufman E, Lamster IB (2002) The diagnostic applications of saliva—a review. *Crit Rev Oral Biol Med* 13(2):197–212 doi: <https://doi.org/10.1177/154411130201300209>
21. Hemdan NYA, Birkenmeier G, Wichmann G, El-Saad AMA, Krieger T, Conrad K, et al. (2010) Interleukin-17-producing T helper cells in autoimmunity. *Autoimmun Rev* 9(11):785–792 doi:10.1016/j.autrev.2010.07.003
22. Hamza YK, Al-Ghurabi BH (2025) Association between the CCR6/CCL20 axis and IL-17A level in patients with rheumatoid arthritis. *World Acad Sci J* 7:65 doi: 10.3892/wasj.2025.353
23. Atwa SE, Azab MM, Mohamed MS (2020) Serum interleukin-17 level in patients with rheumatoid arthritis and its relation to disease activity. *Zagazig Univ Med J* 26(1):87–93 doi: 10.21608/zumj.2019.11577.1196
24. Moran EM, Mullan R, McCormick J, Connolly M, Sullivan O, FitzGerald O, et al. (2009) Human rheumatoid arthritis tissue production of IL-17A drives matrix and cartilage degradation: synergy with tumour necrosis factor- α , oncostatin M and response to biologic therapies. *Arthritis Res Ther* 11:1–12 doi: 10.1186/ar2772
25. Rosu A, Margaritescu CL, Stepan A, Musetescu A, Ene M (2012) IL-17 patterns in synovium, serum and synovial fluid from treatment-naïve, early rheumatoid arthritis patients. *Rom J Morphol Embryol* 53(1):73–80 PMID: 22395503
26. Jain M, Attur M, Furer V, Todd J, Ramirez R, Lock M, et al. (2015) Increased plasma IL-17F levels in rheumatoid arthritis patients are responsive to methotrexate, anti-TNF, and T cell costimulatory modulation. *Inflammation* 38:180–186 doi: 10.1007/s10753-014-0020-1
27. Alturaiqi W, Alhamad A, Alturaiqi M, Mir SA, Iqbal D, Bin Dukhyil AA, et al. (2022) Assessment of IL-1 β , IL-6, TNF- α , IL-8, and CCL5 levels in newly diagnosed Saudi patients with rheumatoid arthritis. *Int J Rheum Dis* 25:1013–1019 doi: <https://doi.org/10.1111/1756-185X.14373>
28. Dhaidan SI (2023) MicroRNA-223 regulates NLRP3 and AIM2 inflammasome activation and inflammatory response in relation to oral manifestation of rheumatoid arthritis. PhD dissertation, Dept Oral Med Coll Dent, Baghdad Univ
29. Shao XT, Feng L, Gu LJ, Wu LJ, Feng TT, Yang YM, Wu NP, Yao HP (2009) Expression of interleukin-18, IL-18BP, and IL-18R in serum, synovial fluid, and synovial tissue in patients with rheumatoid arthritis. *Clin Exp Med* 9:215–221 doi: <https://doi.org/10.3892/br.2024.1775>
30. Broz P, Dixit VM (2016) Inflammasomes: mechanism of assembly, regulation and signalling. *Nat Rev Immunol* 16:407–420 doi: 10.1038/nri.2016.58
31. Abdullah WL, Al-Hashimy AF, Al-Ghurabi BH (2024) Role of salivary caspase-1 and gasdermin D in the pathophysiology of rheumatoid arthritis in relation to salivary pH and flow rate of saliva. *World Acad Sci J* 7(1):7 doi: <https://doi.org/10.3892/wasj.2024.295>