

Original Research

Histological and histochemical characterization of the digestive tract of European catfish (*Silurus glanis* Linnaeus, 1758)

S. Köprücü^{1*}, M. Yaman²

¹ Fisheries Faculty, Fırat University, Elazığ 23119, Turkey

² Veterinary Faculty, Fırat University, Elazığ 23119, Turkey

Abstract: In this study was to examined the morphology, histology and histochemically the digestive tract of European catfish, *Silurus glanis*. Two-four age *S. glanis* obtained from Çelik Lake (Gölbaşı-Adıyaman, Turkey). Six *S. glanis*, 59-71 cm length, 1.5-3.2 kg weight used for in the investigation. The body cavity was opened and samples of digestive tract (esophagus, stomach and intestine) were fixed in neutral buffered 10% formalin and embedded in paraffin. Dewaxed section (5-6 µm) were deparaffinized and stained with haematoxylin and eosin (H&E), Crossman triple, periodic acid-shiff (PAS), alcian blue (AB, pH 1.0 and 2.5) and PAS+AB pH 2.5 technique for different structures. The histological and histochemical structures of all specimens were viewed under a light microscope and microphotograph. The histological structure consists of four layer: tunica mucosa, submucosa, tunica muscularis and tunica serosa. The short esophagus of *S. glanis* have numerous deep longitudinal folds, squamous epithelium with numerous mucous cells which react positively to PAS and AB stains. The muscularis mucosa was organized in longitudinal and circular layers of striated muscular fibers. The stomach is sac-like shaped and its mucosa was formed by simple columnar epithelium with folds. Histologically, the stomach shows three different region: cardia, fundus and pylorus region. The surface epithelium of stomach reacted positively to PAS and AB (pH 1.0 and 2.5). The mucosal surface of intestine of *S. glanis* has numerous folds lined by simple tall columnar cells. There were large number goblet cells in the intestine. The goblet cells reacted to PAS and AB (pH 1.0 and 2.5).

Key words: *Silurus glanis*, histology, histochemical, digestive tract.

Introduction

Fish digestive tracts show remarkable differences in morphology and function. Differences observed at specific levels are related to the food, feeding habits, age, body weight, shape and sex (1-6).

The histology of fish digestive tract has been studied with light and electron microscope in numerous fish species. Generally, the basic histological structures are similar: wall of the digestive tract of many fish is composed of mucosa, submucosa, muscularis and serosa (7-17). The presence of mucosubstances in mucosa of the digestive tract has been observed in most teleosts (18-23).

The mucin layer of wall of digestive tract has various functions such as lubrication, digestion, absorption, protect the tunica mucosa against chemicals, parasites, control of the infectious diseases and colonization of the harmful or opportunistic microorganisms (24-28). Histochemical characterizations of mucins secreted by mucous cells in fish digestive tracts vary among different species, age and regions of the tract (29-32).

Silurus glanis is the largest bodied freshwater fish of Europe. *Silurus* is the only existing genus in Europe of the Siluridae family. It has a triangular-shaped head with small eyes and a large mouth, with two very long, slender. Pigmentation is generally dark along its back with marbled sides, with a greyishwhite. The skin is scale-less, coated in mucus, contains sensory cells (33). *S. glanis* is an economically important species in commercial (34).

Presently, dozens of research projects have been focusing on the geographical distribution, environmental biology, parasitosis and nutriology of this species (33),

feeding behavior (35), embryonic development (36), mucosubstances of the digestive tract (31). However, the information about histology features and mucin histochemistry of its digestive tract has been poorly understood.

The purpose of the present study was to describe histology and histochemically of the digestive tract on the light microscopic level of carnivorous fish species, *S. glanis*.

Materials and Methods

In this study, *S. glanis* obtained from Çelik Lake (Gölbaşı-Adıyaman, Turkey). Six adult *S. glanis* (Length: 59-71 cm, Weight: 1.5-3.2 kg, 2-4 age) were used for in the investigation. Six fish were killed after being anaesthetized in MS-222. The body cavity was opened and samples of digestive tract (esophagus, stomach and intestine) were fixed in neutral buffered 10% formalin and emmedded in paraffin. Dewaxed section (5-6 µm) were deparaffinized and stained for general morphological purpose with haematoxylin and eosin (H&E) (37), Crossman triple stains (38). Periodic acid-shiff (PAS) was used for demonstration of neutral mucosubatans. Alcian blue (AB) at pH 2.5 and pH 1.0 and were used for demonstration of various kinds of acid

Received June 6, 2016; Accepted November 03, 2016; Published November 30, 2016

* Corresponding author: S. Köprücü, Fisheries Faculty, Fırat University, Elazığ 23119, Turkey. Email: skoprucu@firat.edu.tr

Copyright: © 2016 by the C.M.B. Association. All rights reserved.

mucosubstans (37). Microphotography were taken with an light microscope.

Results

The digestive tract of *S. glanis* is composed of the esophagus, stomach and intestine. The short esophagus was followed by a sac-shaped stomach. The intestine had same thickness throughout and convoluted structure. Histologically, the wall of the digestive tract was made up of the mucosa, submucosa, tunica muscularis and tunica serosa.

The esophagus of *S. glanis* was found to have numerous deep longitudinal folds; it was lined by a few layers of stratified squamous epithelium with the numerous superficial mucous cells (Figure 1). All mucous cells show a moderate to high content of PAS-positive stained because of the neutral mucosubstances. But only some of them were positively stained by AB due to the acidic mucosubstance. The lamina propria was formed by the connective tissue but the lamina muscularis mucosa was not present. The muscularis was composed of two striated muscular layers, a thick inner circular and thinner outer longitudinal layer. The serosa consist of the mesothelial cells, small blood vessels, blood cells and loose connective tissue.

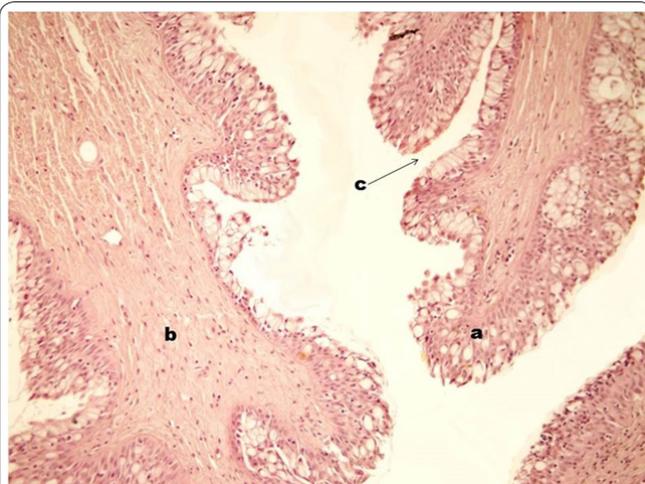


Figure 1. Esophagus; mucus cells (a), lamina propria (b), gastric pit (c) (H&E, x200).

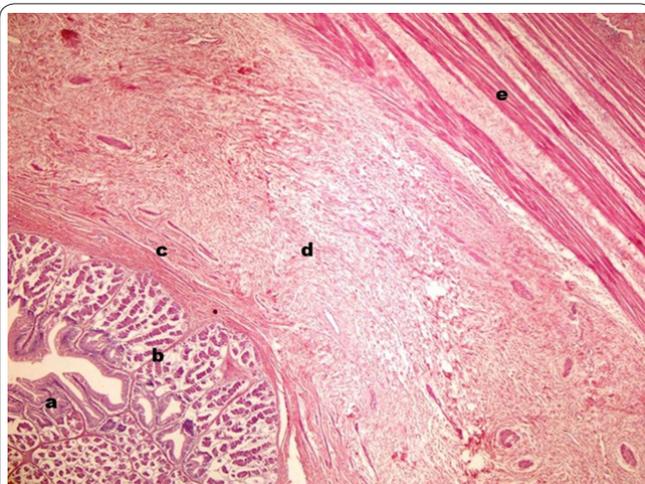


Figure 2. Cardiac stomach; tunica mucosa (a), gastric glands (b), lamina propria (c), submucosa (d), tunica muscularis (e), (H&E, x40).

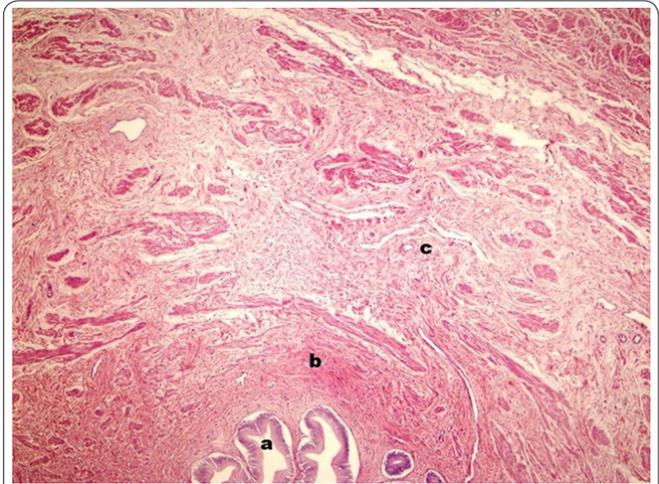


Figure 3. Pyloric stomach; tunica mucosa (a), lamina propria (b), submucosa (c), (H&E, x100).

The stomach of *S. glanis* included the cardiac, fundus and pyloric region. The mucosa of the cardiac, fundus and pyloric were formed with the longitudinal folds that were shallow in the fundus. There were numerous gastric pits of the stomach formed by the invaginations of the mucosal layer into the lamina propria. The epithelium of mucosa was consisted of a single layer of the columnar cells and mucus cells. In the cardiac and fundic stomach, a plenty of gastric glands were observed between the epithelium and lamina propria (Figure 2), whereas in the pyloric stomach the epithelium connected the lamina propria directly and no glands existed (Figure 3). The gastric glands were branched tubular and surrounded by a layer of the connective tissue and opened into the bottom of gastric pits (Figure 4). The glands consisted of a single layer of cells. Mucus cells and the apical cytoplasm of the epithelial cells shows the presence of mucous substances stained strongly with PAS (Figure 5) and AB (pH 1.0 and 2.5). In the staining of PAS+AB pH 2.5 reacted positively to PAS in the apical portion of the cytoplasm and positively to AB in the mucus cells. A small amount of PAS-positively stained substances is observed in the lamina propria. The submucosa is a thin layer having a very vascularized and dense connective tissue. The muscularis was made up of two layers, an inner circular and an outer longitudinal layer of the smooth muscle. Nerve plexus was present between these two muscular layers. The

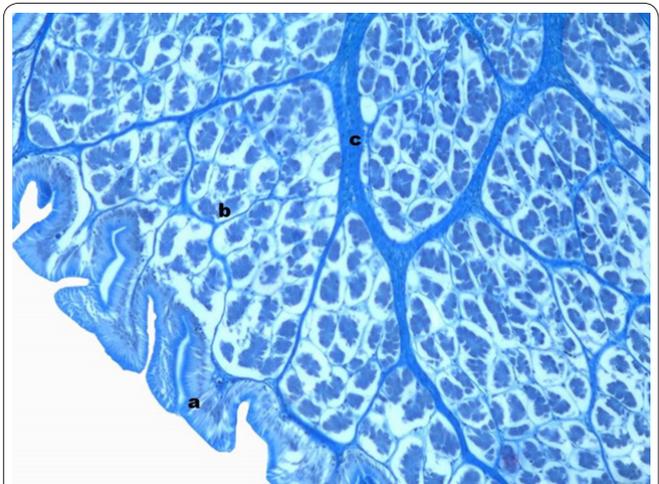


Figure 4. Stomach; columnar epithelial cells (a), gastric glands (b), connective tissue (c), (Crossman, x200).

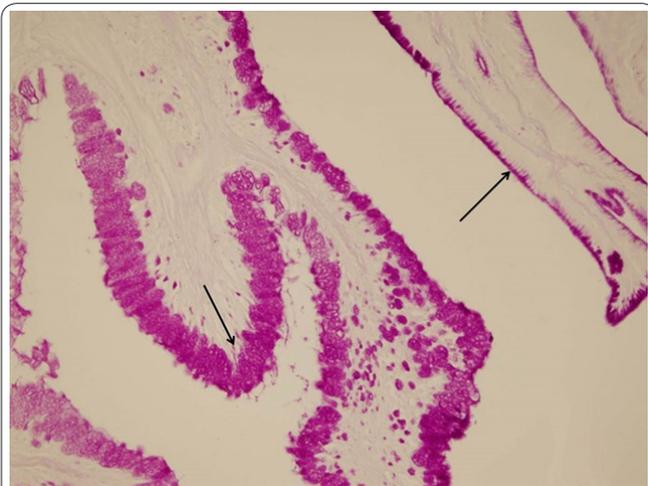


Figure 5. Stomach; PAS positive mucus cells and apical region (PAS, X200).

pyloric stomach had a thickest muscularis among all the parts of digestive tract. The serosa is composed by the connective tissue, mesothelium and blood vessels.

There was no pyloric caeca observed between the stomach and anterior intestine. The intestine is short and its mucosa has an almost uniform structure throughout its entire length. The mucosal surface in the intestine had numerous elongated and deep folds lined. This intestine is lined with simple columnar epithelium which is bordered by a layer of microvilli on their apical surface. The large numbers mucus-secreting goblet cells were found between the epithelial cells as well as occasional leucocytes and macrophages could be seen in the mucosa (Figure 6). The lamina propria-submucosa was formed by loose connective tissues with the many vessels and blood cells. The goblet cell contents stain intensively with AB (pH 1.0 and 2.5) (Figure 7) and they are also PAS-positive (Figure 8). The apical surface of the epithelial cells has a thin PAS-positive and AB (pH 1.0 and 2.5) brush border. The tunica muscularis was organized in two distinct layers of the smooth muscle; the inner circular and outer longitudinal layers. Between both layers can be observed neuronal mienteric plexus.

Discussion

As described in many other fishes (9, 10, 12-14, 16, 17, 39, 40) the wall of digestive tract in the *S. glanis* was

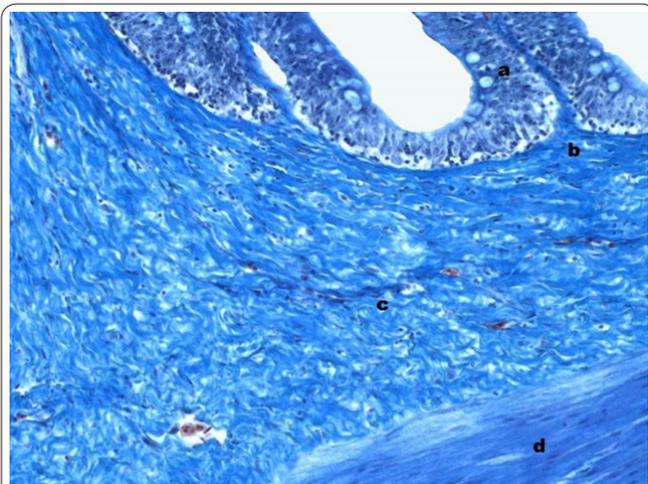


Figure 6. Intestine; tunica mucosa (a), lamina propria (b), submucosa (c), tunica muscularis (d), (Crossman, x200).

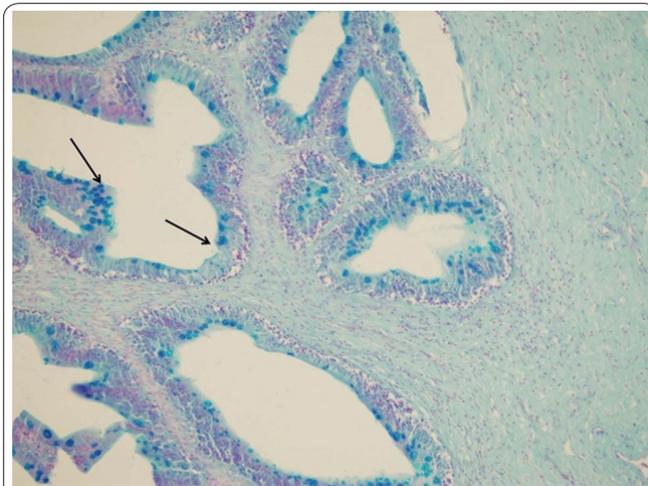


Figure 7. Intestine; AB positive goblet cells (ABX200).

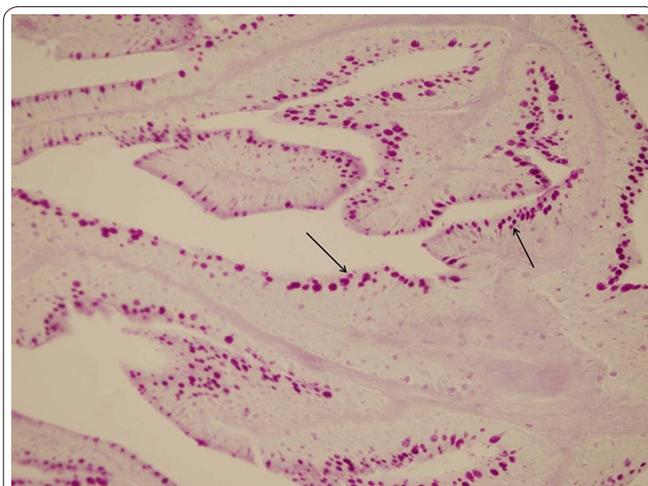


Figure 8. Intestine; PAS positive mucus cells (PASX200).

divided into four layers.

The anatomy of the esophagus of *S. glanis* resembles that of the other teleosts (7, 9, 14, 17, 32, 40). The mucosal folds of the esophagus described by Cataldi et al. (7), Diaz et al. (16) and Hernandez et al. (32) were similar to the esophagus of *S. glanis*. The present study revealed that the esophageal epithelium is stratified squamous with the mucous cells. The mucous cells were mainly PAS positive, indicating that their contents are of the neutral mucosubstans, although some contain AB positive, indicating their contents of acidic mocosubstans. This secretion is similar to the *Acipenser transmontanus* (20), *Rhamdia quelen* (32), *S. glanis* (31, 36), *Tilapia spilurus* and *Mylio cuvieri* (41). Lamina muscularis mucosa were not observed in the wall of the esophagus in the *S. glanis*, which is similar to the *Claris batrachus*, *Serrasalmus nattereri* (40) and *Polyodont spathula* (42). The muscle layers of *S. glanis* had the outer circular and inner longitudinal layers. It was the existence of a thick striated muscle fibril. These result were similar to that described by Simsek and Sarieyyuoglu (9), Suiçmez and Ulus (14), Diaz et al. (16), Hernandez et al. (32), Raji and Norouzi (40).

Generally, the stomach of fish is divided into categories depending on their shape; no stomach, straight with an enlarged lumen, U-shaped, Y-shaped and sac-shaped. The stomach is absent in many fish, including *Crypripnids*, where straight stomachs are rare, and seen in the carnivorous fishes, including in the pike, channel catfish and halibut, while *salmonide* are an example of fish

with a U-shaped stomach (40). The present study revealed that the sac-shaped stomach consist of three regions; the cardiac, fundus and pylorus, which was similar to *Orthrias angorae* (14), *R. quelen* (32), *Pelteobagrus fulvidraco* (39) and *Oreochromis niloticus* (43). The primary and secondary folds of the stomach, described by Cataldi et al. (7), Simsek and Sarieyyupoglu (9) and Grau et al. (29) were similar to the stomach of *S. glanis*.

The gastric glands in *S. glanis* were well-developed in the cardiac and fundic stomach, and absent in the pyloric stomach. The gastric glands consisted of mucous cells are arranged as tubular branched glands surrounded by connective tissue. Mucus cells and the apical borders of the epithelial cells in *S. glanis* shows the presence of mucous substances stained strongly with PAS and AB (pH 1.0 and 2.5). In the staining of PAS+AB pH 2.5 reacted positively to PAS in the apical portion of the epithelial cell and positively to AB in the mucus cells. The acid mucosubstances are shown to be chiefly of the carboxylate and sialylated type. In fish, neutral mucosubstances secreted by the stomach epithelium have been related to the absorption of easily digested molecules, such as disaccharide and short-chain fatty acids (29). The mucosubstances facilitate the movement of large-sized food particles, as well as protecting the gastric mucosa from mechanical injury, chemicals, parasites, control infectious disease (24-28, 31). The presence of both neutral and acid mucosubstances was observed in the superficial gastric epithelium of the *A. transmontanus* (20), *R. quelen* (32), *P. fulvidraco* (39) and in developing larvae and adults of the *Sparus aurata* (19).

There was no pyloric caeca observed between the stomach and anterior intestine of *S. glanis* which is similar to the *P. fulvidraco* (39), *C. batrachus* (40).

In many fish as the *Oncorhynchus mykiss* (9, 17) and *Chanos chanos* (44) the submucosa presents an stratum compactum. That structure were not observed in the present study on the *S. glanis*, which agree with previous studies realized by Hernandez et al. (32) in the *R. quelen*, Sis et al. (45) in the *Ictalurus punctatus* and Pignalleri et al. (46) in the *Pimelodus albicans*. According to the results obtained from present study, the folds in the epithelium of intestine were abundant, the intestine of *S. glanis* contains numerous goblet cells in the mucosa which react positively to PAS and AB (pH 1.0 and 2.5). The presence of mucous-producing goblet cells in the intestinal mucosa has been reported in many fish studied earlier (9, 17-20, 32, 39, 47). The mucous secreted by goblet cells in the intestine has many functions. For example, it lubricates undigested materials for onward progression into the rectum. In addition, It has got a possible role in osmoregulation (31). The studies of Ribelles et al. (48) have shown that the quality of gut mucosubstances is directly related to environmental conditions, which in turn may directly affect the function of the alimentary tract. The presence of mucosubstances, especially those sulfated in the intestine, possibly regulate the transfer of proteins, or a fragment of them, as well as of ions and fluids (19, 24, 31). Like many other fishes (17, 32, 39, 47) a large number of microvilli and blood vessels were observed in the intestine of *S. glanis*.

This investigation, as well as previous investigations on the other fishes, demonstrates that the quality of gut

mucosubstances varies in the different regions of alimentary canals of the *S. glanis*. It appears that histological structure and the quality of mucosubstances in particular parts of the digestive tract is similar to the investigated carnivorous fish. That results can offer a baseline for the future detailed immunohistochemical studies in the *S. glanis*.

Acknowledgements

The fish in this experiment were acted in agreement with the international rules for the conservation of animal welfare.

References

1. Ghosh A, Das KM, Ghosh A. Morphohistology of the digestive tract of a mullet, *Liza parsia* (Ham) in relations to its food habits. J Indian Soc Coast Agric Res 1987; 5:437-444.
2. Boglione C, Bertolin B, Russiello M, Cataudella S. Embryonic and larval development of thicklipped mullet (*Chelon labrosus*) under controlled reproduction conditions. Aquaculture 1992; 101:349-359.
3. Murray HM, Wright GM, Goff GP. A comparative histological and histochemical study of the stomach from three species of Pleuronectid, the Atlantic halibut, *Hippoglossus hippoglossus*, the yellowtail flounder, *Pleuronectes ferruginea*, and the winter flounder, *Pleuronectes americanus*. Can J Zool 1994; 72:1199-1210.
4. Murray HM, Wright GM, Goff GP. A comparative histological and histochemical study of the post-gastric alimentary canal from three species of pleuronectid, the Atlantic halibut, the yellowtail flounder and the winter flounder. J Fish Biol 1996; 48:187-206.
5. Buddington RK, Krogdahl A, Bakke-Mc Kellep AM. The intestines of carnivorous fish: structure and function and the relations with diet. Acta Physiol Scand 1997; 161(Suppl. 638):67-80.
6. Gordon AK, Hecht T. Histological studies on the development of the digestive system of the clownfish *Amphiprion percula* and the time of weaning. J Appl Ichthyol 2002; 18:113-117.
7. Cataldi E, Cataudella S, Monaco G, Rossi A, Tancioni L. A study of the histology and morphology of the digestive tract of the Sea-Bream, *Sparus auratus*. J Fish Biol 1987; 30:135-145.
8. Caceci T, Hrubec TC. Histology and ultrastructure of the gut of black mollie (*Poecilia* spp.), a hybrid Teleostei. J Morphol 1990; 204:265-280.
9. Şimşek S, Sarieyyüpoğlu M. Histological study on digestive tract of the rainbow trout (*Oncorhynchus mykiss*, Walbaum, 1792). Fırat University, Journal of Science and Engineering 1996; 8(1):131-146.
10. Tibbetts IR. The distribution and function of mucous cells and their secretions in the alimentary tract of *Arrhamphus sclerolepis krefftii*. J Fish Biol 1997; 50:809-820.
11. Sarieyyüpoğlu M, Girgin A, Köprücü S. Histological study in the digestive tract on larval development of rainbow trout (*Oncorhynchus mykiss*, Walbaum, 1792). Turk J Zool 2000; 24:199-205.
12. Park JY, Kim JS. Histology and mucin histochemistry of the gastrointestinal tract of the mud loach, in relation to respiration. J Fish Biol 2001; 58:861-872.
13. Park JY, Kim JS, Kim SY. Structure and mucous histochemistry of the intestinal respiratory tract of the mud loach, *Misgurnus anguillicaudatus* (Cantor). J Appl Ichthyol 2003; 19:215-219.
14. Suiçmez M, Ulus E. A study of the anatomy, histology and ultrastructure of the digestive tract of *Orthrias angorae* Steindachner, 1897. Folia Biologica 2005; 53:1-2.
15. Mojazi AB, Bakrkazemi M, Pousti I, Vilaki AS. A histological study on the development of the digestive tract of Caspian salmon, *Salmon trutta caspius* (Kessleri), from hatching to parr stage. Iranian

J Fish Sci 2005; 5:63-84.

16. Diaz AO, Escalante AH, García AM, Goldemberg AL. Histology and histochemistry of the pharyngeal cavity and oesophagus of the silverside *Odontesthes bonariensis* (Cuvier and Valenciennes). *Anat Histol Embryol* 2006; 35:42-46.

17. Khojasteh SMB, Sheikhzadeh F, Mohammadnejad D, Azami A. Histological and ultrastructural study of the intestine of Rainbow trout (*Oncorhynchus mykiss*). *World Appl Sci J* 2009; 6:1525-1531.

18. Nachi AM, Hernandetblasquez FJ, Barbieri RL, Leite RG, Ferri S, Phan MT. Intestinal histology of a detritivorous (iliophagous) fish *Prochilodus scrofa* (Characiformes, Prochilodontidae). *Ann Sci Nat Zool* 1998; 19:81-88.

19. Domeneghini C, Stranini RP, Veggetti A. Gut glycoconjugates in *Sparus aurata* L. (Pisces, Teleostei), a comparative histochemical study in larval and adult ages. *Histol Histopathol* 1998; 13:359-372.

20. Domeneghini C, Arrighi S, Radaelli G, Bosi G, Mascarello GS. Morphological and histochemical peculiarities of the gut in white sturgeon, *Acipenser transmontanus*. *European J Histochemistry* 1999; 43:135-145.

21. Pedini V, Scocco P, Radaelli G, Fagioli O, Ceccarelli P. Carbohydrate histochemistry of the alimentary canal of the Shi Drum, *Umbrina Cirrosa* L. *Anat Histol Embryol* 2001; 30:345-349.

22. Domeneghini C, Radaelli G, Bosi G, Arrighi S, Di-Giancamillo A, Pazzaglia M, Mascarello F. Morphological and histochemical differences in the structure of alimentary canal in feeding and runt (feed deprived) white sturgeon (*Acipenser transmontanus*). *J Appl Ichthyol* 2002; 18:341-346.

23. Bosi G, Arriqhi S, Di-Giancamillo A, Domeneghini C. Histochemistry of glycoconjugates in Mucous cells of *Salmo trutta* uninfected and naturally parasitized with intestinal helminths. *Dis Aquat Org* 2005; 64:45-51.

24. Gupta BJ. The relationship of mucoid substances and ion and water transport, with new data on intestinal goblet cells and a model for gastric secretion. *Symp Soc Exp Biol* 1989; 43:81-110.

25. Zaccone G, Fasulo S, Ainis L, Contini A. Localization of calmodulin positive immunoreactivity in the surface epidermis of the brown trout, *Salmo trutta*. *Histochemie* 1989; 91:13-16.

26. Loretz, CA. Electrophysiology of ion transport in teleost intestinal cells. In: Cellular and Molecular Approaches to Fish Ionic Regulation. Wood CH. and Shuttleworth TJ. (eds.), *Academic Press, London*, 1995, pp. 25-56.

27. Veggetti A, Rowlerson A, Radaelli G, Arrighi S, Domeneghini C. Post-hatching development of the gut and lateral muscle in the sole, *Solea solea* (L). *J Fish Biol* 1999; 55 (Suppl. A):44-65.

28. Domeneghini C, Arrighia S, Radaelli G, Bosia G, Veggetti A. Histochemical analysis of glycoconjugate secretion in the alimentary canal of *Anguilla anguilla* L. *Acta Histochem* 2005; 106:477-487.

29. Grau A, Crespo S, Sarasquete MC, Gonzales De Canales ML. The digestive tract of the amberjack *Seriola dumerii*, Risso: a light and scanning electron microscope study. *J Fish Biol* 1992; 41:287-303.

30. Sarasquete C, Gisbert E, Ribeiro L, Vieira L, Dinis MT. Glycoconjugates in epidermal, branchial and digestive mucous cells and gastric glands of gilthead sea bream, *Sparus aurata*, Senegal sole, *Solea senegalensis* and Siberian sturgeon, *Acipenser baeri* development. *Eur J Histochem* 2001; 45:267-278.

31. Petrinc Z, Nejedly S, Kuzir S, Opacak A. Mucosubstances of

the digestive tract mucosa in northern pike (*Esox lucius* L.) and European catfish (*Silurus glanis* L.). *Vet Archiv* 2005; 75:317-327.

32. Hernandez DR, Perez Ganeselli M, Domitrovic HA. Morphology, histology and histochemistry of the digestive system of South American Catfish (*Rhamdia quelen*). *Int J Morphol* 2009; 27(1):105-111.

33. Gordon HC, Robert BJ, Cucherousset J, Garcia-Berthou E, Kirk R, Peeler E, Stakenas S. Voracious invader or benign feline? A review of the environmental biology of European catfish in its *Silurus glanis* native and introduced ranges. *Fish and Fisheries* 2009; 10:252-282.

34. Adamek Z, Fasaic K, Siddiqui MA. Prey selectivity in wels (*Silurus glanis*) and African catfish (*Clarias gariepinus*). *Ribarstvo* 1999; 57:47-60.

35. Omarov OP, Popova OA. Feeding behavior of pike, *Esox lucius* and catfish, *Silurus glanis*, in the Arakum Reservoirs of Dagestan. *J Ichthyol* 1985; 25:25-36.

36. Kozaric Z, Kuzir S, Petrinc Z, Gjurcovic E, Bozic M. The development of the digestive tract in larval European catfish (*Silurus glanis* L). *Anat Histol Embryol* 2008; 37:141-146.

37. Luna LG. *Manual of Histology Staining Methods*. McGraw-Hill Book Co., New York, 1968, pp. 72-100.

38. Crossman G. Modification of mallory's connective tissue stain with a discussion of the principles involved. *Anat Rec* 1937; 69:33-38.

39. Cao XJ, Wang WM. Histology and mucin histochemistry of the digestive tract of yellow catfish, *Pelteobagrus fulvidraco*. *Anatomia, Histologia, Embryologia* 2009; 38(4):254-261.

40. Raji AR, Norouzi E. Histological and histochemical study on the alimentary canal in Walking catfish (*Claris batrachus*) and piranha (*Serrasalmus nattereri*). *Iran J Vet Res* 2010; 11(32):255-261.

41. Abdulhadi HA. Some comparative histological studies on alimentary tract of Tilapia fish (*Tilapia spilurus*) and sea bream (*Mylio cuvieri*). *Egypt J Aquat Res* 2005; 31:387-397.

42. Weisel GF. Anatomy and histology of the digestive system of the paddlefish (*Polyodon spathula*). *J Morphol* 2005; 140:243-255.

43. Caceci T, El-habback HA, Smith SA, Smith BJ. The stomach of *Oreochromis niloticus* has three regions. *J Fish Biol* 1997; 50:939-952.

44. Ferraris RP, Tan JD, De La Cruz MC. Development of the digestive tract of milkfish, *Chanos chanos* (Forsskal): histology and histochemistry. *Aquaculture* 1987; 61:241-257.

45. Sis RE, Ives RJ, Jones DM, Lewis DH, Haensly WE. The microscopic anatomy of the oesophagus, stomach and intestine of the channel catfish, *Ictalurus punctatus*. *J Fish Biol* 1979; 14:179-86.

46. Pignalberi C, Cordiviola de Yuan E, Occhi R. Anatomia e histologia del aparato digestivo de *Pimelodus albicans* (Valenciennes) (Pises, Pimelodidae). *Phycis Secc. B. Buenos Aires* 1973; 32:297-308.

47. Rodrigues APO, Pauletti P, Kindlein L, Cyrino JEP, Delgado EF, Machado-Neto R. Intestinal morphology and histology of the striped catfish *Pseudoplatystoma fasciatum* (Linnaeus, 1766) fed dry diets. *Aquacult Nutr* 2009; 15(6):559-563.

48. Ribelles A, Carrasco MC, Rosety M, Aldana M. A histochemical study of the biological effects of sodium dodecyl sulfate on the intestine of the gilthead seabream, *Sparus aurata* L. *Ecotox Environ Safe* 1995; 32:131-138.