

**Table S1.** Primers Used to Amplified Exon 1-5 of *AR* Gene.

Exon	Forward Primer (5' – 3')	Reverse Primer (5' – 3')	Amplicon Size
1.A	AAGATTCAGCCAAGCTCAAG	GATACTGCTTCCTGCTG CTG	629
1.B	GCTGACCTTAAAGACATCCTG	CAGAGCCAGTGGAAAGTTG	579
2	GCAGGTAAATGCTGAAGACC	ACCTTCACTGCCTAAATTGC	458
5	CACTGTCACCCCATCACC	TAGCTCAACCCGTCAGTACC	285
7	TCAGATCGGATCCAGCTATC	TTGGCTCTATCAGGCTGTTC	417
8	AGGTTGGGGAAGAGGCTAGC	GCACTGCAGAGGAGTAGTGC	320

**Table S2.** Master Mixture for Four Different Ready-To-Use PCR Kits.

PCR Kit	Mixture
MyTaq HS Red Mix 2x (BioLine)	12,5 µl MyTaq HS Red Mix 2x 1µl (10µM) of each primers Mili Q water up to 25 µl
FastStart Taq DNA Polymerase, dNTPack (Roche) (standard mixture)	2,5 µl PCR reaction buffer 10x concentration with 20mM MgCl <sub>2</sub> 5 µl GC-Rich Soution 5x concentration 0,5 µl (10mM) dNTP solution 0,2 µl (5U/µl) Taq DNA Polymerase 1µl (10µM) of each primers Mili Q water up to 25 µl
FastStart Taq DNA Polymerase, dNTPack (Roche) (2xTaq, 2x dNTPs mixture)	2,5 µl PCR reaction buffer 10x concentration with 20mM MgCl <sub>2</sub> 5 µl GC-Rich Soution 5x concentration 1 µl (10mM) dNTP solution 0,5 µl (5U/µl) Taq DNA Polymerase 1µl (10µM) of each primers Mili Q water up to 25 µl
KAPA2G fast PCR Kit 2X (Kapa Biosystem)	12,5 µl 2X KAPA2G Fast ReadyMix (with 1,5 mM MgCl <sub>2</sub> ) 1µl (10µM) of each primers Mili Q water up to 25 µl
KOD FX Neo (Toyobo)	12,5 µl 2x PCR Buffer for KOD FX Neo 5 µl (2mM) dNTPs 1µl (1U/µl) KOD FX Neo 1µl (10µM) of each primers Mili Q water up to 25 µl

**Table S3.** Touch-Down PCR Program.

	Temperature	Duration	Annotation
Initial PCR	95°C	5 minutes	
Denaturation	95°C	1 minutes	
Annealing	68°C *	1 minutes	*Lower annealing temperature by 1°C at every cycle for the first 10 cycles to 59°C
Extension	72°C	1 minutes	
Denaturation	95°C	1 minutes	
Annealing	58°C#	1 minutes	#Repeated for 20 cycles once annealing temperature reach 58°C
Extension	72°C	1 minutes	
Final Extension	72°C	5 minutes	