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Biological activities and health-promoting effects of *Pyracantha* genus: a key approach to the phytochemical's potential

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Abstract: Pyracantha spp. are commonly called firethorn, and attract human attention due to their colorful berries. These berries are eaten globally as a traditional remedy for treating different stomach abnormalities, and as a cooking ingredient for folk diets. The present review aims to provide an overview on Pyracantha genus' geographical distribution and botanical description, traditional uses, phytochemical composition, biological activities and safety issues. Several biological activities have been reported to Pyracantha species, namely antioxidant, anti-inflammatory, antimicrobial, larvicidal and cytotoxic properties, most of them attributed to the use of their fruits. Pyracantha species phytochemical composition reveal the presence of interesting bioactive molecules, such as pyracrenic acid and fortuneanosides. The currently reported biological activities to these plants derive from in vitro and in vivo studies, so that clinical trials are needed to confirm these preclinical results. Nonetheless, Pyracantha species can be suggested as a safe herb useful to develop future drug formulations and functional foods.

Key words: Pyracantha genus; Pyracrenic acid; Fortuneanoside; Phytochemicals; Biological effects.

Introduction

Pyracantha genus is an evergreen, thorny shrub belonging to the Rosaceae family, widely used in gardening and landscaping (1). The genus is commonly called as 'firethorn' due to the presence of dense poisonous thorns when pierce the human skin causes severe pain and inflammation. Originally introduced and planted as garden ornamentals, Pyracantha genus appears to house about 10 species (2-5). However, the exact number of species is unclear as there are many synonyms inconsistently used in the literature. Because of aesthetic values and presence of thorns, these plants are popularly used for making fences and walls.

In this sense, the present review aims to provide an overview on *Pyracantha* spp geographical distribution, botanical description, traditional uses, phytochemical

composition, biological activities and safety issues.

Geographical distribution

Pyracantha spp. prefer warm temperate (Mediterranean) to cool, subtropical climates, with many species preferring cool subtropical climates (6-8). Pyracantha spp. are native to an area extending from Southwest Europe east to Southeast Asia. Various Pyracantha species are broadly distributed from the Eastern Mediterranean region and central Europe across Asia to China and Taiwan (9) (Table 1).

Botanical description

Plants under *Pyracantha* genus are thick shrub to small tree (1-4 m tall), occasionally reaching up to 6 m

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Table 1. Species, common names and Distribution of *Pyracantha* species.

Pyracantha species	Common name	Distribution	Ref.
P. angustifolia (Franch.) C. K. Schneid.	Narrow leaf firethorn, Slender firethorn and Woolly firethorn	Chinese provinces of Guizhou, Hubei, Sichuan, Xizang, Yunnan and Zhejiang; central Argentina	(62, 63)
P. atalantioides (Hance) Stapf	Gigg's firethorn and Sichuan firethorn	Southeastern and western China	(62)
P. coccinea M. Roem.	Scarlet firethorn	Temperate Asia and Europe (Iran, Lebanon, Turkey, Armenia, Azerbaijan, Georgia, Ukraine, Albania, Bulgaria, Former Yugoslavia, Greece, France, Spain, central and southern Italy)	(30)
P. crenatoserrata (Hance) Rehder	Huo ji (China)	Chinese provinces of Fujian, Guangxi, Guizhou, Henan, Hubei, Hunan, Jiangsu, Shaanxi, Sichuan, Xizang, Yunnan and Zhejiang	(10)
P. crenulata (Roxb. ex D. Don) M. Roem.	Himalayan firethorn, Chhota seb,Nepalese firethorn, Indian hawthorn, "Ghingharu"	Temperate Himalayas; Chinese provinces; Bhutan and India	(20, 64-66)
P. densiflora T. T. Yu		China	(10)
P. fortuneana (Maxim.) H. L. Li	Broadleaf firethorn	Central China	(10)
P. inermis J. E. Vidal		China	(10)
P. koidzumii (Hayata) Rehder	Red berry firethorn, Formosa firethorn	Taiwan	(10, 26)
P. rogersiana (A. B. Jacks.) ColtmRog.	Asian firethorn	Western China	(10)

in height and spreading up to 5 m across. Stem is erect to divergent or bent at a sharp angle, grayish bark in color. Plants are usually with long and short branches, glabrous or tormentous. Thorns are present mostly on short branches. Leaves are sessile or shortly petiolate, stipulate, simple, alternate, lanceolate to ovate up to 5 cm long, dark green, pubescent, margin crenulate, serrulate, or entire; venation camptodromous. Inflorescence terminal, compound corymb, many flowered; peduncle sub-glabrous to pubescent. Flowers are white to creamy white, blooms in spring and summer, pentamerous, pedicellate, bracteate, bract caduceus, lanceolate; bracteoles present; sepals 5, triangular, persistent, abaxially pubescent when young, apex acute; petals 5, ovate to sub-orbicular to broadly elliptic, white to creamy white; stamens 15-20, epigynous, filament 2-4 mm, shorter than petals, anthers yellow, dorsifixed, bilocular; hypanthium campanulate; carpels 5, syncarpous, ovary pubescent, 5 locular, ovules 2 in each locule; placentation axile; style free, nearly equal to stamen. Fruit fleshy pome, looks like tiny apple, 5-9 mm in diameter, red-to-orange color, sometimes yellow, glabrous. Seeds are brown and irregular in shape (4, 10).

Traditional uses

Plants of *Pyracantha* genus are used in traditional cultures in countries where they were originally endemic. They have been used in China for more than 1700 years as a food source, and have also been used as food in the Western Himalayas. At present, fruit juice, fruit wine and preserved fruit from these plants remain popular (11, 12).

Pyracantha fortuneana (Maxim.) H. L. Li is used as skin-whitening agent in cosmetics in Japan (13), and Pyracantha crenulata (Roxb. ex D. Don) M. Roem. is a fuel in Himalayan countries (11). Nowadays, plants of Pyracantha genus are popular in countries with a Mediterranean climate as a landscape ornamental (14).

In Ayurvedic medicine, *Pyracantha* spp. is used in the treatment of a number of disorders, including hepatic, stomach and skin diseases (15).

In traditional herbal medicine in some cultures, 4 species of Pyracantha genus are used. In the traditional medicine of the Indian Himalayas, P. crenulata fruits are often prescribed for the treatment of hypertension, arteriosclerosis, angina, diabetes, pregnant anemia, stomach disorders, including constipation, and externally in the form of juice for the treatment of earache (16-19). In traditional medicine of Nepal (20) and Indian rural community (21), the fruits of this plant are eaten in cases of dysentery. In Uttarakhand, bark of this plant is recommended for heavy bleeding during menstrual cycles (22). Also leaf paste of *P. crenulata* is used for burns (23). In Central Himalaya, P. crenulata roots are used in bathing for body pain (24). In traditional Chinese Medicine, P. fortuneana plants are used as a popular remedy. Fruits of this plant are recommended for oral administration for the treatment of dyspepsia and dysentery or for externally for sores relief (13, 25). Pyracantha koidzumii (Hayata) Rehder is used in dysmenorrhea (26). In Europe, P. coccinea fruits are taken for medicinal use as heart soother (27). Leaves of this plant are used in diarrhea and urinary diseases in Turkey (27, 28). P. coccinea are also used in veterinary medicine for diarrhea (27).

Phytochemical composition

Several studies showed the presence of phytochemicals, like glycosides, carbohydrates, steroids, flavonoids, phenolics, resins and tannins in *Pyracantha* spp. (18, 29-31). For example, Fico, A (32) found phenolic acids, such as hyperoside, isoquercitrin, quercetin and rutin in P. coccinea fruits. They also found that the flavonoids profile of P. coccinea showed marked differences between aerial and hypogeal parts of the plant. P. fortuneana fruits were also known to contain several types of polyphenols, such as cyanidins (33), biphenyl glycosides (34, 35), fortuneanosides A to F (34-36), acylphloroglucinol glycosides (34), and dibenzofuran glycosides derivatives (34, 35). The ripe fruit of Pyracantha angustifolia (Franch.) C. K. Schneid. constituted the best practical source for the isolation of pro-y-carotene and has been a good source for prolycopene (37). A new compound, pyracrenic acid, isolated from P. crenulata bark revealed to be a potent anti-inflammatory agent (38). Quiroga, Bou (39) reported that linoleic (61.1%), oleic (17.3%) and palmitic (17.4 %) acids were the major fatty acids present in *P. crenulata* seed extract. Two compounds, viz. 3,6-dihydroxy-2,4-dimethoxy-dibenzofuran and 3,4-dihydroxy-5-methoxybiphenyl-2'-O-D-glucopyranoside were isolated from P. koidzumii (26). Overall, in the genus *Pyracantha*'s fruiting and flowering parts, around 30 metabolites could be found. Pyracanthoside and rutin, two flavonoids, appear during all stages of the plant life cycle (32). Representative phytochemicals present in *Pyracantha* spp. are shown in Figure 1.

Biological activities

Several bioactive effects have been reported to *Pyracantha* spp., with the most prominent ones being their antioxidant, antimicrobial, larvicidal, cytotoxic, and anti-inflammatory effects. In the following sections are described the biological activities of the most common *Pyracantha* spp., summarized in Figure 2.

Pyracantha angustifolia (Franch.) C.K.Schneid.

Kim, Park (40) reported that *P. angustifolia* leaves methanol extract (at 10, 000 ppm) exhibited insecticidal and acaricidal activities against Tetranychus urticae, Aphis gossypii, Myzus persicae, Trialeurodes vaporariorum, and Panonychus citri. The antioxidant activity, total polyphenols and electron donating abilities of methanol extract, methylene chloride, ethyl acetate, and methanol fractions of *P. angustifolia* were reported by Lee (29). The extract exhibited superoxide radical as well as hydrogen peroxide (H₂O₂) scavenging activity. The highest total polyphenol contents (2007.30±109.28 μg GAE/mL) were found in 70% methanol extract. Moreover, 70% methanol extract also showed highest electron donating abilities (79.07±7.31) and superoxide radical removal ability (0.018±0.003). Furthermore, the methylene chloride fraction exhibited maximum H₂O₂ decomposition (0.0027±0.0015) among the extracts/ fractions.

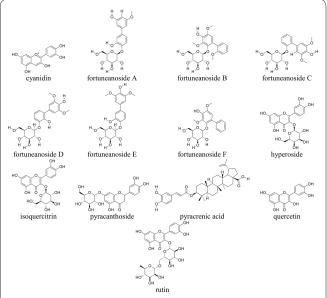
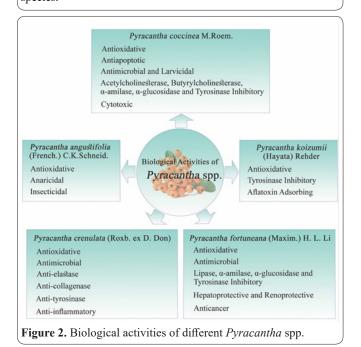


Figure 1. Representative phytochemicals present in *Pyracantha* species.



Pyracantha coccinea M. Roem.

Antioxidant activity of P. coccinea was reported by several authors (9, 30, 41). Min, Yeon (41) reported that P. coccinea root extract (5 µg/ml), among all extracts tested [leaf, stalk and root for 6 days (1, 5 and 10 µg/ ml respectively)] significantly increased blastocyst formation, total cell numbers and reduced apoptotic index in porcine parthenogenetic embryos. It decreased intracellular levels of reactive oxygen species (ROS) under H₂O₂-induced oxidative stress. Therefore, the improvement in the *in vitro* development of porcine parthenogenetic embryos was attributed to the Pyracantha root extract via its antioxidant and antiapoptotic properties. In other study, Keser (9) used different solvents to extract phytochemicals from *P. coccinea* fruit and determined antiradical activities by 2,2-diphenyl-1-picrylhydrazyl (DPPH), 2,2'-Azino-bis(3-ethylbenzothiazoline-6-sulfonic acid) (ABTS) and hydroxyl radical scavenging assays. Ethanol, methanol and acetone extracts exhibited higher scavenging activity on ABTS when compared to

BHT, used as positive control, whereas methanol and ethanol extracts showed higher scavenging activity on DPPH compared to BHT. The extracts also showed better antioxidant activities than α - and δ -tocopherol, vitamins K and D, β-sitosterol, ergosterol, and stigmasterol. Sarikurkcu and Tepe (30) reported that the ethanol and water extracts of P. coccinea inhibited acetylcholinesterase, butyrylcholinesterase, α-amylase, α-glucosidase and tyrosinase effects. It was also observed that the scavenging potential on DPPH radical, ABTS radical cation, superoxide anion and nitric oxide was higher for the ethanol extract than that of water extract. Ethanol extracts also displayed higher activities in antioxidant power (FRAP), cupric ion reducing, ferric reducing, iron (III) to iron (II) reduction, and phosphomolybdenum assays. The work suggested its possible therapeutic use against Alzheimer's disease.

To P. coccinea extracts, interesting antimicrobial (27, 42) and larvicidal (43) effects have been also reported. Turker, Yildirim (27) reported that *P. coccinea* extracts (from 8 solvents) exhibited antibacterial activity against Staphylococcus aureus, Staphylococcus epidermidis and Streptococcus pyogenes. Among the extracts, fruits extracted in hot ethanol exhibited prominent antibacterial activities against the 3 tested bacteria. Kokubun, Harborne (42) reported that *P. coccinea* sapwood-derived dibenzofuran phytoalexin, 9-hydroxyeriobofuran exhibited antifungal activity against Alternaria alternata, Botrytis cinerea and Fusarium culmorum, comparable to that of cotonefurans. 9-hydroxyeriobofuran (10, 20, 50 and 100 µg/ml) inhibited fungal spore germination with, respectively, 100% 100% and 98% inhibition (ED₅₀ values of 44, 39 and 49 μ g/ml), at 10 μ g/ml, against the 3 fungi. Koc, Evren (43) reported that P. coccinea fruit methanol extract (62.5-1000 ppm) exerted larvicidal activity against second instar larvae of Culex pipiens. At 62.5 ppm, following 96 h of exposure, the extract showed larvicidal activity (0.46±0.26%) with LC_{50} and LC_{90} values of 32110.12 and 808652.32 ppm, respectively.

 $P.\ coccinea$ fruits methanolic extract also revealed cytotoxic effects Vahabi, Monajemi (44). In that study, it was reported a weak cytotoxic effect (with highest activity at 800 µg/ml) on Hela cell lines (MTT assay). The antioxidant activity (DPPH assay), and total phenolic contents (Folin-Ciocalteu assay) displayed by the methanol extract were found to be higher when compared to the aqueous extract.

Pyracantha crenulata (Roxb. ex D. Don) M. Roem.

Several studies have reported the antioxidant activity of *P. crenulata* (45-47). Pal, Arun Kumar (46) used methanol extract (0.5, 1.0, 1.5 and 2.0 mg/ml) of *P. crenulata* mature fruits and reported radical scavenging activities. In addition, *P. crenulata* possessed higher levels of lycopene and β-carotene when compared to the another studied plant *Berberis asiatica*. Saklani, Badhani (47) reported the antioxidant activity of *P. crenulata* fruits through DPPH [19.18±0.14 ascorbic acid equivalent (AAE)/100 g fw], ABTS (4.41±0.03 AAE/100 g fw) and ferric ion reducing antioxidant power (FRAP) (2.34±0.01 AAE/100 g fw) assays. Besides, it exhibited total phenols, total flavonoids, total anthocyanins, vi-

tamin C and gallic acid at the levels of 5.59±0.05 mg GAE/g fw, 5.46±0.04 mg/g fw, 0.44±0.05 mg/100 g fw, 3.30±0.34 mg/100 g fw and 10.15±1.40 mg/100 g fw, respectively. Singh, Lily (45) reported that *P. crenulata* fruit 80% aqueous acetone extract exhibit free radical scavenging activities (through DPPH, ABTS, linoleate hydroperoxyl and superoxide radicals), FRAP, ferrous metal chelating capacity, anti-elastase, anti-collagenase, anti-tyrosinase and anti-inflammatory activities. The total phenolics and total flavonoids content found were, respectively, 144.16±3.50 mg GAE/100 g fw and 303.75±3.50 mg CE/100 g fw. Otsuka, Fujioka (38) isolated pyracrenic acid from P. crenulata bark and found that it displayed anti-inflammatory activity (in the fertile egg method) at doses of 10, 200 and 400 µg/disc, with inhibition percentage of 13.8%, 38% and 43.1%, respectively, and survival ratios of embryos with 19/20, 16/20, 18/20, respectively. In addition, it also inhibited granulation tissue formation by the fertile egg method, at 50 mg/kg, when orally administered in rats.

Saklani and Chandra (18) reported that *P. crenulata* fruit pulp ethanolic extract exhibit antimicrobial activities against *Shigella flexneri*, *Escherichia coli* and *S. pyogenes*. The crude protein, crude fiber and carbohydrates contents in the extract were found to be 5.13%, 7.40% and 24.88%, respectively. The extract was also reported to contain phenolics, saponins and flavonoids at 1.83%, 1.56% and 3.12%, respectively.

Pyracantha fortuneana (Maxim.) H. L. Li

Diverse biological activities have been reported to P. fortuneana. Zhao, Lei (48) reported the antioxidant activity of *P. fortuneana* fruit water-soluble polymeric polyphenolic fractions on HepG(2) cells. The authors also reported an increase in cell antioxidant activities using peroxyl radical-induced DCFH oxidation method with higher decrement being stated at EC₅₀ values of 2.91. The in vivo antioxidant benefit of small molecular polyphenols was attributed to the additive effects conferred by the proanthocyanidins present in fractions. In another study, the same group also assessed the total polyphenolic content (TPC), total antioxidant activity (TAA) and chromatographic profiling of the plant extract through response surface methodology (RSM) and solvent optimization, in order to enhance the P. fortuneana fruits applicability as nutraceutical in food industries (49).

Wang, Ye (31) with the help of high-pressure liquid chromatography coupled with quadrupole time-of-flight mass spectrometry (HPLC-QTOF-MS/MS) profiling identified 7 flavonoids, 6 phenolic acids, 7 organic acids, 3 tannins, 1 terpene, and 1 alkaloid from *P. fortuneana* fruit (50% acetone extract) and reported antioxidant and α -glucosidase inhibitory activities. Wei, Chai (50) extracted proanthocyanidins from *P. fortuneana* fruit and reported α -glucosidase inhibition (IC₅₀ 0.15 µg/mL) in a non-competitive manner. Dai, Zhou (35) reported the isolation of biphenyl glycosides, fortuneanosides A to E from *P. fortuneana* fruit and observed that fortuneanoside D inhibited tyrosinase (IC₅₀=0.07 mM) most efficiently when compared to arbutin (IC₅₀=0.23 mM). Xu, Zhao (51) reported that *P. fortuneana* fruit extract reduced lactulose/mannitol ratio and glucose transporter 2

(GLUT2), improved tight junction proteins, modulated bacterial groups linked with gut barrier integrity, such as *Actinobacteria*, *Bacteroidaceae*, *Corynebacteriaceae*, *Lactobacillaceae*, and S24-7, and inhibited α-amylase, α-glucosidase, and lipase enzymes. Moreover, the same group reported, in rodents, a decrease in body weight, triacylglycerol, total cholesterol (TC), low-density lipoprotein cholesterol (LDL-c), and malondialdehyde (MDA) levels, and elevation in high-density lipoprotein cholesterol (HDL-c), glutathione peroxidase (GPx), and superoxide dismutase (SOD) (52). These results markedly suggested the *P. fortuneana* fruit potential against hyperlipidemia, obesity and oxidative stress.

Peng, Guo (25) reported that selenium-enriched P. fortuneana polysaccharides (Se-PFPs) exhibit antioxidant and hepatoprotective effects. In mouse liver, at 3 concentrations (1.35, 2.7 and 5.4 g/kg body weight) it was stated a significant inhibition on cyclophosphamide-induced micronucleus formation in both bone marrow and peripheral blood, cytochrome P450 1A and increased SOD and GPx activities. In the same way, Yuan, Li (53) reported that Se-PFPs decreased alanine aminotransferase (ALT), aspartate aminotransferase (AST), LDH, TC, triglycerides in serum, while elevated the SOD, GPx, glutathione and catalase activities. The same group also reported that Se-PFPs inhibited the triple-negative breast cancer, MDA-MB-231 cells growth in a dose-dependent manner at G2 phase, increased p53, B-cell lymphoma-2 (Bcl-2)-associated X Protein (Bax), Puma and Noxa, Bax/Bcl2 ratio, caspases 3/9, and decreased Bcl2. More interestingly, Se-PFPs was reported as containing carbohydrates [93.7% (w/w)], uronic acid [2.1% (w/w)] and selenium $[3.7 \mu\text{g/g}]$. (54). Similarly, Sun, Dong (55) reported that Se-PFPs reduced cyclin D1, Bcl-2 and matrix metallopeptidase 9 (MMP-9), Ncadherin, vimentin, ZEB1 and ZEB2 and increased the cleavage of PARP, caspase-3, and -9, E-cadherin and cytokeratin 19 in HEY and SKOV3 human ovarian cancer cells. The authors have proposed Se-PFPs as promising against ovarian cancer.

Ke, Yu (56) reported that *P. fortuneana* fruit extract (administered orally at 250 mg/kg) protected against the acute renal toxicity induced by cadmium chloride (CdCl₂) (6.5 mg/kg for 5 days) in rats. Critical parameters, like kidney weight, body weight, malondialdehyde (MDA), nitric oxide production, uric acid, urea, and creatinine, glutathione, SOD, catalase, GPx, Bcl-2 expression, Bax, TNF-α, Keap-1, Nrf2, HO-1, γ-GCS, GPx and NQO1 expression were also assessed. Sun, Wang (13) optimized the polyphenols extraction (by ultrasonic assisted extraction method) from P. fortuneana fruits and reported their antibacterial activity against S. aureus and E. coli (MIC values of 10 and 20 mg/mL, respectively). Mei, Wan (57) reported that P. fortuneana extract and the chloroform phase (among the 3 polar extract fractions), when administered orally, shortened the coagulation time of glass slide in mice. In addition, they also shortened the prothrombin time and recalcification time in vitroMei, Wan (57).

Pyracantha koidzumii (Hayata) Rehder

Jiang, Chang (58) reported that *P. koidzumii* extracts exhibited less cytotoxicity than other natural products

used in cosmetics, displayed promising cellular tyrosinase inhibitory activity (IC₅₀=54.8 µg/mL) and potent free radical scavenging activity against hydroxyl, superoxide, and ABTS anion radicals. P. koidzumii leaves and fruits contained, respectively, 3.55 and 0.23 g GAE/g extract of total phenolic contents. Lin, Chen (26) isolated 3,6-dihydroxy-2,4-dimethoxy-dibenzofuran and 3,4-dihydroxy-5-methoxybiphenyl-2'-O-D-glucopyranoside from P. koidzumii and observed that these compounds inhibited cellular tyrosinase in a dose-dependent manner. The compound 3,4-dihydroxy-5-methoxybiphenyl-2'-O-D-glucopyranoside exerted uncompetitive mixed-type inhibitor effects (against L-3,4-dihydroxyphenylalanine) with a K_m value of 0.262 mM. Ramales-Valderrama, Vázquez-Dürán (59) reported that P. koidzumii biomasses (leaves, berries and leaves/berries mixture) adsorbed B-aflatoxins (AFB1 and AFB2) through electrostatic interaction. The highest aflatoxin uptakes were reported to be 86% and 82% at 6 h, respectively, for leaves and leaves/berries mixture biomasses. Hence, P. koidzumii biomasses-mediated biosorption could be viewed as an effective alternative strategy to remove Baflatoxins.

Safety and adverse effects

Pyracantha spp. attracts human attention due to its colorful berries and those are eaten globally as traditional medicine and as a cooking ingredient. However, there is no in-depth report on their safety concerns and adverse effects. P. crenulata fruits are eaten by Indian rural community in several regions, with its powder being also mixed with yoghurt to treat bloody dysentery, thus clarifying its non-toxic properties (21). P. angustifolia leaf extract showed lesser effects while checking its insecticidal activities against A. gossypii, M. persicae, T. vaporariorum, and P. citri, revealing a higher survival rate after 72 h exposure at 10,000 ppm when compared to the control and other plants tested (40). In another study, P. coccinea fruit methanol extract was applied in mosquito larvae (C. pipiens) to determine its larvicidal/insecticidal effects, but no significant mortality was found even at the highest concentration (i.e. 1000 ppm) (43). Pretreatment with *P. fortuneana* fruit extract on kidney injury in CdCl₂-injected rats showed that it could significantly reduce CdCl2-induced toxicities through increasing Nrf2, HO-1, y-GCS, GSH-Px and NQO-1 expression (56). Thus, it was reported to be beneficial for treating renal toxicity caused by CdCl₂. This extract was also reported to have ameliorative effects on intestinal barrier dysfunction in high fat dietinduced Sprague Dawley rats without showing toxicity (52). Besides, P. fortuneana polysaccharides were found to enhance immune function and oxidative stress in mice (60). Of them, selenium containing polysaccharides could significantly reduce the tumor growth besides to confer preventive effects against CCl₄-induce liver injury in mice model without altering body weight (53, 54). Peng, Guo (25) reported that selenium containing polysaccharides administered to mice at doses up to 5.4g/kg body weight caused no detectable toxicity and death during the 30-day experiments. Medicinal plant polysaccharides are in general thought to be nontoxic, thus making them suitable for therapeutic applications (61), and based on the several previous findings, it is undoubtedly clear that selenium containing polysaccharides from *Pyracantha* spp. can be used and considered as a safe naturally-derived therapeutic agent.

All previous studies with *Pyracantha* spp. evidenced its safety and non-toxicity as a traditional medicinal source. However, there are some rumors among rural/scientific communities on the possible plant toxicities. Thus, *Pyracantha* spp. can be suggested as a safe herbal source for developing future drug formulations and functional foods for human health promotion.

Conclusions and future perspectives

Pyracantha spp. are used in folk medicine for treating different human medical conditions, such as stomach abnormalities, hepatic and skin diseases. In addition, several biological properties have been attributed to the Pyracantha genus members, viz. antioxidant, anti-inflammatory, antimicrobial, larvicidal and cytotoxic effects. Many reports are available on specific Pyracantha spp. for their pharmacological benefits using various in vitro and in vivo animal models, most of them concentrated on the effects of its fruits. The use of these plants in folk medicine and its reported biological activities (in vitro and in vivo) are closely related to the plants' phytochemical composition. However, a scarce amount of studies is available elucidating some interesting phytochemicals found in *Pyracantha* spp. (i.e. pyracrenic acid and fortuneanosides). In addition, some Pyracantha spp. extracts and phytochemicals have revealed enzymatic inhibition abilities, and thus may be useful in pathologies such as hyperlipidemia, obesity and oxidative stress-related diseases. Thus, given the current insights, further studies are needed to determine the phytochemical composition of all plant parts of different Pyracantha spp. and to determine the biological activities of plants extracts and purified phytochemicals. Clinical trials should also be conducted to confirm the preclinical studies findings. Anyway, *Pyracantha* spp. can be viewed as safe plants, useful to develop future drug formulations and functional foods.

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Conflicts of Interest

The authors declare no conflict of interest.

References

- 1. Novarra L. Rosaceae. Flora del Valle Lerma. Aportes Botánicos Salta, Flora. 1993;2:14-144.
- 2. Meyer FG, Mazzeo PM, Voss DH. A Catalog of Cultivated Woody Plants of the Southeastern United States. USDA Agricultural Research Service, U.S. National Arboretum Contr. No. 7. U.S. Govt. Printing Office, Washington. 1994.
- 3. Egolf DR, Andrick AO. A Checklist of Pyracantha Cultivars. U.S. National Arbor. Contr. No. 8. U.S. Govt. Printing Office, Washington. 1995.

- 4. Gu C, Spongberg SA. Pyracantha. Flora of China. 2003;9:108-
- 5. The Plant List 2020 [Available from: http://www.theplantlist.org/.6. Xiwen L, Walker D. The Plant Geography of Yunnan Province, Southwest China. Journal of Biogeography. 1986;13(5):367-97.
- 7. Wang W, Wang Q, Li S, Wang G. Distribution and Species Diversity of PlantCommunities along Transect on the Northeastern Tibetan Plateau. Biodiversity & Conservation. 2006;15(5):1811-28. 8. Liu C-C, Liu Y-G, Guo K, Zheng Y-R, Li G-Q, Yu L-F, et al. Influence of drought intensity on the response of six woody karst species subjected to successive cycles of drought and rewatering. Physiologia Plantarum. 2010;139(1):39-54.
- 9. Keser S. Antiradical activities and phytochemical compounds of firethorn (Pyracantha coccinea) fruit extracts. Natural Product Research. 2014;28(20):1789-94.
- 10.Csurhes S, Markula A. Invasive plant risk assessment: Firethorn Pyracantha species. Bitou bush Department of Agriculture and Fisheries Biosecurity Qld. 2016.
- 11.Uniyal S, Awasthi A. Developmental Processes, Changing Lifestyle and Traditional Wisdom: Analyses from Western Himalaya. The Environmentalist. 2003;23:307-12.
- 12. Wang Y, Li Y, Liu J, Liu L, Yang Q, Deng Q. Analysis of nutritional components of the fruits of Pyracantha angustifolia and *Pyracantha fortunaeana*. IOP Conference Series: Earth and Environmental Science. 2018;199:052013.
- 13.Sun H, Wang X, Zhou Z, Wang R. Extraction optimization of polyphenols from fruits of *Pyracantha fortuneana* (Maxim.) Li by ultrasonic assistant method and their antibacterial activity. Pakistan Journal of Pharmaceutical Sciences. 2019;32(4):1635-41.
- 14. Hummer K, Janick J. Rosaceae: Taxonomy, Economic Importance, Genomics. Genetics and Genomics of Rosaceae, Plant Genetics and Genomics: Crops and Models. 2009;6:1-17.
- 15.Bahuguna YM, Rawat MSM, Juyal V, Gusain K. Evaluation of *Pyracantha crenulata* Roem for antiurolithogenic activity in albino rats. African Journal of Urology. 2009;15(3):159-66.
- 16.Parihaar R, Bargali K, Bargali S. Diversity and uses of Ethnomedicinal plants associated with traditional agroforestry systems in Kumaun Himalaya. Indian Journal of Agricultural Sciences. 2014;84(12):1470-6.
- 17.Singh H, Agnihotri P, Pande P, Husain T. Biodiversity conservation through a traditional beliefs system in Indian Himalaya: A case study from Nakuleshwar sacred grove. Environmentalist. 2011;31:246-53.
- 18.Saklani S, Chandra S. In vitro antimicrobial activity, nutritional value, antinutritional value and Phytochemical screening of *pyracantha crenulata* fruit. International Journal of Pharmaceutical Sciences Review and Research. 2014;26(1):1-5.
- 19. Arya KR. Traditional uses of some common plants in indigenous folklore of Dronagiri: mythic hill of Uttarakhand. Indian Journal of Traditional Knoweledge. 2002;1(1):81-6.
- 20.Kunwar RM, Shrestha KP, Bussmann RW. Traditional herbal medicine in far-west Nepal: a pharmacological appraisal. Journal of ethnobiology and ethnomedicine. 2010;6:35.
- 21. Arora RK, Pandey A. Wild edible plants of India: diversity, conservation and use: National Bureau of Plant Genetic Resources New Delhi; 1996.
- 22.Sharma IP, Kanta C, Chandra S, Goswami N. Wild Fruits of Uttarakhand (India): Ethnobotanical and Medicinal Uses. International Journal of Complementary & Alternative Medicine. 2017;8(3):1-8. 23.Bisht VK, Kandari LS, Negi JS, Bhandari AK, Sundriyal RC. Traditional use of medicinal plants in district Chamoli, Uttarakhand, India. Journal of Medicinal Plants Research. 2013;7(15):918-29. 24.Negi C, Nautiyal S, Dasila L, Rao K, Maikhuri R. Ethnomedicinal Plant Uses in a Small Tribal Community in a Part of Central

- Himalaya, India. Journal of human ecology. 2002;14(1):23-31.
- 25.Peng F, Guo X, Li Z, Li C, Wang C, Lv W, et al. Antimutagenic Effects of Selenium-Enriched Polysaccharides from *Pyracantha* fortuneana through Suppression of Cytochrome P450 1A Subfamily in the Mouse Liver. Molecules. 2016;21(12).
- 26.Lin RD, Chen MC, Liu YL, Lin YT, Lu MK, Hsu FL, et al. New whitening constituents from taiwan-native Pyracantha koidzumii: Structures and tyrosinase inhibitory analysis in human epidermalmelanocytes. International Journal of Molecular Sciences. 2015;16(12):28598-613.
- 27.Turker AU, Yildirim AB, Karakas FP. Antibacterial and antitumor activities of some wild fruits grown in Turkey. Biotechnology & Biotechnological Equipment. 2012;26(1):2765-72.
- 28.Dogan A, Bulut G, Senkardes I, Tuzlacı E. An ethnopharmacological analysis of Rosaceae taxa in Turkey. The 2016 WEI International Academic Conference Proceedings (Boston, USA). 2016:44-51
- 29.Lee K-S. A Study of Antioxidant Effects of *Pyracantha angustifolia* (Franch.) C. K. Schneid Extract. The Korean Journal of Food and Nutrition. 2017;30(6):1286-91.
- 30.Sarikurkcu C, Tepe B. Biological activity and phytochemistry of firethorn (*Pyracantha coccinea* MJ Roemer). Journal of Functional Foods. 2015;19:669-75.
- 31. Wang H, Ye Y-h, Wang H-h, Liu J, Liu Y-j, Jiang B-W. HPLC-QTOF-MS/MS profiling, antioxidant, and alpha-glucosidase inhibitory activities of *Pyracantha fortuneana* fruit extracts. Journal of Food Biochemistry. 2019;43(5).
- 32.Fico G, A RB, Morelli II, Tome F. Flavonoid distribution in Pyracantha coccinea plants at different growth phases. Biochemical systematics and ecology. 2000;28(7):673-8.
- 33.Ma B-L, Liang S-F, Xie Y-F. A study on separation and stability of cyanidin from *Pyracantha fortuneana*. Journal of Northwest Forestry University. 2006;21:162-5.
- 34.Dai Y, Zhou G-X, Kurihara H, Ye W-C, Yao X-S. Fortuneanosides G-L, dibenzofuran glycosides from the fruit of *Pyracantha* fortuneana. Chemical & Pharmaceutical Bulletin. 2008;56(4):439-42.
- 35.Dai Y, Zhou GX, Kurihara H, Ye WC, Yao XS. Biphenyl glycosides from the fruit of Pyracantha fortuneana. Journal of Natural Products. 2006;69(7):1022-4.
- 36.Dai Y, Zhou G-X, Kurihara H, Ye W-C, Yao X-S. A biphenyl glycoside from *Pyracantha fortuneana*. Natural Product Research. 2009;23(13):1163-7.
- 37.Zechmeister L, Schroeder WA. The fruit of Pyracantha angustifolia: a practical source of pro- γ -carotene and prolycopene. Journal of Biological Chemistry. 1942;144(2):315-20.
- 38.Otsuka H, Fujioka S, Komiya T, Goto M, Hiramatsu Y, Fujimura H. Studies on Anti-inflammatory Agents. V. A New Anti-inflammatory Constituent of *Pyracantha crenulata* Roem. Chemical and Pharmaceutical Bulletin. 1981;29(11):3099-104.
- 39. Quiroga OE, Bou SM, Sarlingo CI, Nolasco SM. Study of the composition of *Pyracantha crenulata* Roem seed, oil and meal. Grasas Y Aceites. 2003;54(4):335-8.
- 40.Kim DI, Park JD, Kim SG, Kuk H, Jang MS, Kim SS. Screening of Some Crude Plant Extracts for Their Acaricidal and Insecticidal Efficacies. Journal of Asia-Pacific Entomology. 2005;8(1):93-100.
- 41.Min S, Yeon J, Kim J, Park S, Lee Y, Kang S, et al. Pyracantha Extract Acts as an Antioxidant Agent to Support Porcine Parthenogenetic Embryo Development In Vitro. Korean Journal of Embryo Transfer. 2013;28(3):243-50.
- 42.Kokubun T, Harborne JB, Eagles J, Waterman PG. Dibenzofuran phytoalexins from the sapwood tissue of Photinia, *pyracantha* and Crataegus species. Phytochemistry. 1995;39(5):1033-7.
- 43.Koc S, Evren OH, Cetin H. Evaluation of Some Plant Fruit

- Extracts for the Control of West Nile Virus Vector Culex pipiens (Diptera: Culicidae). Journal of Arthropod-Borne Diseases. 2016;10(4):595-601.
- 44. Vahabi L, Monajemi R, Hosseini SA. The cytotoxic effect of methanolic extract of Pyracanthacoccinea M. Roemer fruit on hela cell line, antioxidant capacities and total phenol contents of methanolic and aquatic extract of this fruit. Biomedical and Pharmacology Journal. 2015;8:99-103.
- 45.Singh H, Lily MK, Dangwal K. Evaluation and comparison of polyphenols and bioactivities of wild edible fruits of North-West Himalaya, India. Asian Pacific Journal of Tropical Disease. 2015;5(11):888-93.
- 46.Pal RS, Arun Kumar R, Agrawal PK, Bhatt JC. Antioxidant capacity and related phytochemicals analysis of methanolic extract of two wild edible fruits from North Western Indian Himalaya. International Journal of Pharma and Bio Sciences. 2013;4(2):P113-P23.
- 47.Saklani S, Badhani A, Mishra AP, Chandra S. Health Promoting Phytochemicals Their Concentration and Antioxidant Activity of Wild Edible Fruits of Uttarakhand, India. Asian Journal of Chemistry. 2012;24(12):5558-60.
- 48.Zhao CF, Lei DJ, Song GH, Zhang H, Xu H, Yu LJ. Characterisation of water-soluble proanthocyanidins of *Pyracantha fortuneana* fruit and their improvement in cell bioavailable antioxidant activity of quercetin. Food Chemistry. 2015;169:484-91.
- 49.Zhao CF, Li S, Li SJ, Song GH, Yu LJ, Zhang H. Extraction optimization approach to improve accessibility of functional fraction based on combination of total polyphenol, chromatographic profiling and antioxidant activity evaluation: *Pyracantha fortuneana* fruit as an example. Journal of Functional Foods. 2013;5(2):715-28.
- 50.Wei M, Chai WM, Yang Q, Wang R, Peng Y. Novel Insights into the Inhibitory Effect and Mechanism of Proanthocyanidins from Pyracantha fortuneana Fruit on α -Glucosidase. Journal of Food Science. 2017;82(10):2260-8.
- 51.Xu H, Zhao C, Li Y, Liu R, Ao M, Li F, et al. The ameliorative effect of the *Pyracantha fortuneana* (Maxim.) H. L. Li extract on intestinal barrier dysfunction through modulating glycolipid digestion and gut microbiota in high fat diet-fed rats. Food & Function. 2019;10(10):6517-32.
- 52.Xu H, Zhao CF, Tian GG, Qian LR, Yu LJ. Characterization of active constituents in *Pyracantha fortuneana* fruit extract and their effects on hyperlipidaemia, obesity, and oxidative stress in rodents. Journal of Functional Foods. 2016;22:278-90.
- 53. Yuan C, Li Z, Yi M, Wang X, Peng F, Xiao F, et al. Effects of Polysaccharides from Selenium-enriched Pyracantha fortuneana on Mice Liver Injury. Medicinal Chemistry. 2015;11(8):780-8.
- 54. Yuan C, Wang C, Wang J, Kumar V, Anwar F, Xiao F, et al. Inhibition on the growth of human MDA-MB-231 breast cancer cells in vitro and tumor growth in a mouse xenograft model by Se-containing polysaccharides from *Pyracantha fortuneana*. Nutrition Research. 2016;36(11):1243-54.
- 55.Sun Q, Dong M, Wang Z, Wang C, Sheng D, Li Z, et al. Selenium-enriched polysaccharides from *Pyracantha fortuneana* (Se-PFPs) inhibit the growth and invasive potential of ovarian cancer cells through inhibiting β -catenin signaling. Oncotarget. 2016;7(19):28369-83.
- 56.Ke Y, Yu K, Zeng W, Lian G. Protective roles of Pyracantha fortuneana extract on acute renal toxicity induced by cadmium chloride in rats. Acta Cirurgica Brasileira. 2019;34(7).
- 57.Mei X, Wan G, Zhou Z, Chang J, Wu H. Effects of *Pyracantha fortuneana* extract on blood coagulation. Zhong yao cai = Zhongyaocai = Journal of Chinese medicinal materials. 2001;24(12):874-6. 58.Jiang CB, Chang MJ, Wen CL, Lin YP, Hsu FL, Lee MH. Natural products of cosmetics: Analysis of extracts of plants endemic to Taiwan for the presence of tyrosinase-inhibitory, melanin-reducing, and

free radical scavenging activities. Journal of Food and Drug Analysis. 2006;14(4):346-52.

- 59.Ramales-Valderrama R, Vázquez-Durán A, Méndez-Albores A. Biosorption of B-aflatoxins using biomasses obtained from formosa firethorn [Pyracantha koidzumii (Hayata) Rehder]. Toxins. 2016;8(7).
- 60.Yuan C, Wang C, Bu Y, Xiang T, Huang X, Wang Z, et al. Antioxidative and immunoprotective effects of Pyracantha fortuneana (Maxim.) Li polysaccharides in mice. Immunology Letters. 2010;133(1):14-8.
- 61.Xie J-H, Jin M-L, Morris GA, Zha X-Q, Chen H-Q, Yi Y, et al. Advances on Bioactive Polysaccharides from Medicinal Plants. Critical reviews in food science and nutrition. 2016;56(sup1):S60-S84.

- 62. Delucchi G. Especies adventicias y naturalizadas nuevas o críticas para Argentina. Boletin de la Sociedad Argentina de Botánica. 1991;27:179-81.
- 63.Morrone O, Zuloaga FO. Catálogo de las plantas vasculares de la República Argentina. [St. Louis]: Missouri Botanical Garden; 1996. 64.Brandis D. Indian Trees: an account of trees, shrubs, woody climber, bamboos & palms, indigenous or commonly cultivated in the British Indian empire. Constable & Company, Ltd, London. 1921:294.
- 65. Weber E. Invasive plant species of the world: a reference guide to environmental weeds: CABI Publishing, United Kingdom; 2003. 66. Singh R, Negi PS, Dwivedi SK. Indian Hawthorn (Pyracantha crenulata). In: Singh B, Peter K, editors. New Age Herbals. Springer, Singapore 2018. p. 135-49.