

Original Article

Clinical significance of miR-200a in systemic lupus erythematosus and renal damage in children



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Abstract



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The purpose was to analyze the clinical significance of miR-200a in children with initially diagnosed SLE and renal damage. Children with initially diagnosed SLE (n=100) and healthy children (n=50) undergoing physical examinations during the same period were recruited. Disease activity of SLE children was determined based on SLEDAI (systemic lupus erythematosus disease activity index), and they were divided into SLEDAI \leq 9 group and SLEDAI>9 group, respectively. Moreover, SLE children were divided into LN and non-LN groups based on the occurrence of lupus nephritis. Differential level of miR-200a between groups was detected by qRT-PCR. Spearman correlation test was conducted to analyze the influence of miR-200a on SLEDAI and other laboratory indexes of SLE children. Its diagnostic potential in SLE and LN was assessed through depicting ROC curves. MiR-200a level was remarkably lower in SLE children than that of healthy children. Lower level of miR-200a was detected in SLE children with high SLEDAI or accompanied LN. MiR-200a level was negatively correlated to SLEDAI

I ($r=-0.425$), ESR ($r=-0.284$), CRP ($r=-0.338$), BUN ($r=-0.263$) and Scr ($r=-0.345$), while it was positively correlated to C3 ($r=0.631$), C4 ($r=0.524$) and ALB ($r=0.394$) in SLE children. The AUC of miR-200a in diagnosing SLE was 0.8379 (cut-off value=2.225, sensitivity=70%, specificity=70%). Besides, the AUC of miR-200a in diagnosing LN was 0.7619 (cut-off value=2.005, sensitivity=80%, specificity=76%). MiR-200a level has a certain correlation to the disease activity of children with initially diagnosed SLE, which can be utilized as an adjuvant indicator in evaluating SLE severity. Meanwhile, miR-200a has predictive value for SLE-induced renal damage.

Keywords: SLE, LN, miR-200a, Diagnosis

1. Introduction

Systemic lupus erythematosus (SLE) is an autoimmune disease, which can involve multiple systems and organs. The main pathological change is vasculitis mediated by the immune complex. SLE is characterized by severe interferences in the functions and activities of innate and adaptive immune cells. Disease manifestations, pathological processes, and clinical outcomes vary significantly among individuals, ethnicities, and age groups. About 10-20% of SLE cases initiate in childhood or puberty, which mainly affects the population in 12-16 years [1]. Compared with adult patients, SLE children have more atypical early manifestations, more dangerous progression, more rapid involvement of organs and worse prognosis [2].

Lupus nephritis (LN) is an important complication of SLE, with diverse clinical manifestations, including asymptomatic hematuria and/or proteinuria, nephrotic syndrome and acute progressive nephritis with renal dysfunction. About 40%-70% of SLE children have clinical manifestations of LN, and the incidence of LN in SLE children is 10-30% higher than in SLE adults. It is reported that 90% of SLE patients suffer from renal damage as renal biopsy results suggest. Seriously, 5-20% of LN patients will aggravate uremia within 10 years [3]. Recently, the incidence of childhood SLE is on the rise, as well as that of LN [4].

MiRNAs are a type of endogenous, non-coding, single-stranded RNAs, and they have about 25 nucleotides in length [5]. They guide mRNA degradation or post-transcriptionally suppress protein translation by binding 3'UTR of target genes, thereby participating in life activities [6]. Due to the stability, disease specificity, and availability, plasma miRNAs have gradually been discovered as disease biomarkers [7,8]. Plasma miRNAs are capable of diagnosing tumors, determining therapeutic efficacy, and monitoring tumor recurrence and metastasis [9-11]. Their vital regulations in autoimmune diseases have been recognized as well [12,13]. MiR-200a is differentially expressed in tumor samples, which is closely linked to malignant phenotypes of tumor cells and clinical prognosis [14,15]. This study aims to explore the clinical significance of miR-200a in childhood SLE and LN.

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2. Materilas and Methods

2.1. Subjects

This study was approved by the ethics committee of Zhanjiang Central People's Hospital. Signed written informed consent were obtained from the patients and/or guardians. Children with initially diagnosed SLE (n=100) in our hospital were recruited. Inclusion criteria: (1) Age < 18 years; (2) They were diagnosed as SLE based on the standard released by the Systemic Lupus International Collaborating [16]; (3) Clinical data were complete; (4) Other diseases were excluded, including drug-induced lupus, rheumatoid arthritis, blood system diseases, mixed connective tissue diseases, etc. Based on the diagnosis of SLE, lupus nephritis (LN) with any of the following manifestations of renal involvement could be diagnosed: (1) Urinary protein test met any of the following: Positive urine protein qualitative examination three times in one week; Or 24-hour urine protein >150 mg; Or urine protein/urinary creatinine > 0.2 mg/mg; Or higher microalbuminuria than the normal three times a week; (2) > 5 erythrocytes per high power field of vision in centrifugal urine; (3) Abnormal function of glomerulus and/or renal tubules; (4) Abnormal findings in renal biopsy that was consistent with pathological changes of LN. During the same period, fifty healthy children undergoing physical examinations were recruited as a control group.

2.2. Acquisition of clinical data

The following data of each subject were recorded. (1) Routine blood test data: WBC, HGB, PLT, NE, LY, MPV and RDW; (2) Baseline characteristics: Age, sex and clinical manifestations; (3) Laboratory indexes: ALB, BUN, Scr, C3, C4, CRP, ESR, ds-DNA and SLEDAI.

2.3. SLEDAI scoring

The disease activity of SLE children was determined using SLEDAI-2000. According to the symptoms and examinations within 10 days, SLEDAI was assessed with the involvement of 24 clinical indexes on 9 organs and systems. Its total score was 105 grades (<4: no disease activity; 5-9: mild stable disease; 10-14: moderate disease activity; ≥15: active disease) [17].

2.4. qRT-PCR

Serum miRNAs were isolated and reversely transcribed to cDNAs. A qRT-PCR mixture involving 1.0 µl of cDNA, 0.8 µl of primers, 10 µl of 2 × miRNA qPCR Mix and 8.2 µl of ddH₂O was prepared. It was subjected

to qRT-PCR at 95°C for 10 min, followed by 45 cycles at 95°C for 10 s, 60°C for 20 s and 72°C for 12 s. U6 (forward: 5'-CTCGCTTCGGCAGCACATTTT-3' and reverse: 5'-AACGCTTCACGAATTTGCGT-3') was the internal reference. MiR-200a sequences were: Forward: 5'-TAACACTGTCTGGTAACGATGT-3' and reverse: 5'-ATCGTTACCAGACAGTGTATT-3'.

2.5. Statistical analysis

Statistic Package for Social Science (SPSS) 22.0 (IBM, Armonk, NY, USA) was used for statistical analysis. Normally distributed quantitative data were expressed as mean ± standard deviation, and differences between groups were compared by the *t*-test. Enumeration data were expressed as a percentage (%) and compared by the χ^2 test. Spearman correlation test was conducted to analyze the influence of miR-200a on SLEDAI and other laboratory indexes of SLE children. The diagnostic potential of miR-200a was assessed by depicting ROC curves. *P*<0.05 was considered as statistically significant.

3. Results

3.1. Comparison between SLE children and healthy subjects

No significant differences in age and sex rate were detected between SLE children and healthy subjects (*P*>0.05). Lower levels of WBC, HGB, PLT and LY, and higher levels of RDW and MPV were detected in SLE children than in healthy subjects. In addition, the serum level of miR-200a was lower in SLE children compared with that of healthy subjects (Table 1). MiR-200a may be involved in the progression of SLE.

3.2. Correlation between miR-200a and laboratory indexes relevant to SLE activity

The recruited 100 SLE children were divided into SLEDAI≤9 group (n=41) and SLEDAI>9 group (n=59). Higher levels of ESR and CRP, as well as higher rate of positive anti-dsDNA, were detected in SLEDAI>9 group in comparison to the other group. Besides, C3, C4 and miR-200a levels were lower in SLEDAI>9 group (Table 2). It is indicated that miR-200a could affect the disease activity of SLE.

3.3. Correlation between miR-200a and laboratory indexes relevant to SLE-induced LN

Recruited 100 SLE children were divided into LN group (n=68) and non-LN group (n=32). Higher levels of

Table 1. Comparison of clinical data between SLE group and control group.

Variable	SLE group (n=100)	Control (n=50)	<i>t</i> / χ^2	<i>P</i>
Age (years old)	10.4±2.1	10.6±2.8	-0.490	0.625
Male/Female	21/79	8/42	0.534	0.518
WBC (×10 ⁹ /L)	4.1±1.15	6.2±1.34	-9.969	<0.001
HGB (g/L)	90±12.1	128±17.3	-15.630	<0.001
PLT (×10 ⁹ /L)	130±25.3	247±31.7	-24.489	<0.001
NE (×10 ⁹ /L)	2.4±0.75	2.51±0.81	-0.824	0.411
LY (×10 ⁹ /L)	1.5±0.09	2.3±0.21	-32.644	<0.001
RDW (%)	15.8±2.4	12.9±2.0	7.358	<0.001
MPV (fl)	8.7±1.02	8.5±0.92	1.169	0.244
miR-200a	1.31±0.56	2.41±0.97	-8.796	<0.001

Table 2. Comparison of laboratory indicators between high and low SLEDAI groups.

Variable	SLEDAI≤9 (n=41)	SLEDAI>9 (n=59)	t/ χ^2	P
C3 (g/L)	0.35±0.07	0.31±0.05	3.335	0.001
C4 (g/L)	0.05±0.03	0.03±0.01	4.763	<0.001
ESR (mm/h)	32±8.22	47±11.53	-7.157	<0.001
CRP (mg/L)	1.14±0.88	2.31±0.93	-6.324	<0.001
Anti-ds-DNA antibody (Positive/Negative)	18/23	39/20	4.864	0.040
miR-200a	2.72±1.04	2.14±0.85	3.060	0.003

Table 3. Comparison of laboratory indexes between LN group and non-LN group.

Variable	LN group (n=68)	non-LN group (n=32)	t	P
ALB (g/L)	30.13±7.42	34.64±8.03	-2.762	0.007
BUN (mmol/L)	5.2±1.42	4.5±1.17	2.426	0.017
Scr (μmmol/L)	53.15±19.74	44.82±16.39	2.073	0.041
SLEDAI	15±2.15	10±1.06	12.439	<0.001
miR-200a	2.14±0.86	2.77±1.13	-3.081	0.003

Table 4. Correlation analysis of miR-200a and various indicators.

Variable	r	P
SLEDAI	-0.425	0.002
ESR (mm/h)	-0.284	0.018
CRP (mg/L)	-0.338	0.007
C3 (g/L)	0.631	<0.001
C4 (g/L)	0.524	<0.001
BUN (mmol/L)	-0.263	0.006
Scr (μmmol/L)	-0.345	0.021
ALB (g/L)	0.394	0.028

BUN, Scr and SLEDAI, as well as lower levels of ALB and miR-200a were detected in LN group in comparison to non-LN group (Table 3). It is suggested that low serum level of miR-200a may trigger the incidence of LN in SLE children.

3.4. Correlation between miR-200a and laboratory indexes relevant to SLE activity

Spearman correlation test was conducted to analyze the influence of miR-200a on SLEDAI and other laboratory indexes of SLE children. MiR-200a level was negatively correlated to SLEDAI ($r=-0.425$), ESR ($r=-0.284$), CRP ($r=-0.338$), BUN ($r=-0.263$) and Scr ($r=-0.345$), while it was positively correlated to C3 ($r=0.631$), C4 ($r=0.524$) and ALB ($r=0.394$) in SLE children (Table 4).

3.5. Diagnostic potential of miR-200a in SLE and LN

To ascertain the prognostic potential of miR-200a in SLE and renal damage, ROC curves were depicted. The AUC of miR-200a in diagnosing SLE was 0.8379 (cut-off value=2.225, sensitivity=70%, specificity=70%) (Figure 1A). Besides, the AUC of miR-200a in diagnosing LN was 0.7619 (cut-off value=2.005, sensitivity=80%, specificity=76%) (Figure 1B).

4. Discussion

Childhood SLE is a chronic autoimmune disease characterized by vascular inflammation and connective tissue inflammation. Positive expressions of specific antinuclear antibodies and anti-dsDNA can be detected in SLE pa-

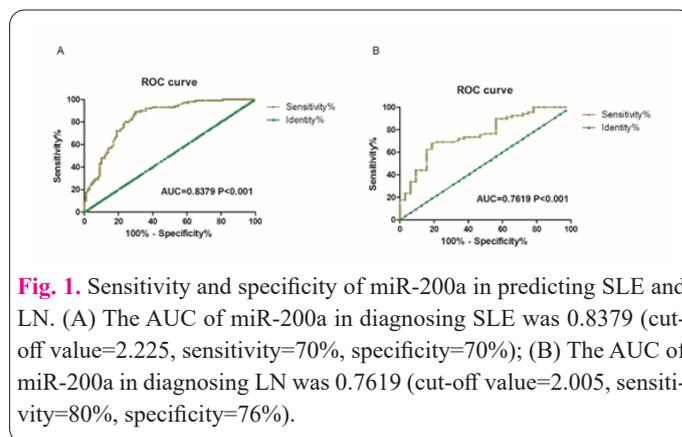


Fig. 1. Sensitivity and specificity of miR-200a in predicting SLE and LN. (A) The AUC of miR-200a in diagnosing SLE was 0.8379 (cut-off value=2.225, sensitivity=70%, specificity=70%); (B) The AUC of miR-200a in diagnosing LN was 0.7619 (cut-off value=2.005, sensitivity=80%, specificity=76%).

tients [18]. The incidence of childhood SLE varies in different races, ranging from 10/100,000-20/100,000, and it covers 15-50% of SLE cases. The 10-year survival of SLE is nearly 90%. However, the life quality of SLE children is poor [19]. About 67-82% of SLE children suffer from renal damage, mainly manifested as proteinuria and hematuria [20,21].

During the active phase of SLE, a large number of immune complexes are deposited in tissues and organs, which activate the complement system to eliminate them through complements-induced classical and bypass pathways [22,23]. As a result, serum C3 and C4 are abundantly consumed. CRP is an acute phase reaction protein and it can activate the complement system. CRP level in the remission phase of SLE decreases significantly compared to the active phase, and it is positively correlated with

SLEDAI [22]. Recent evidences have proven the close relation between red blood cell distribution width and autoimmune diseases. RDW is an independent risk factor for autoimmune hepatitis (AIH)-induced cirrhosis, which is a promising indicator for revealing the progression of AIH [24]. Tecer D et al. [25] suggested that RDW is able to reflect inflammatory state of rheumatoid arthritis, and it is linked to the disease activity and pain degree. Platelet activation is an inflammatory marker. Platelet activation in SLE patients may be related to immune complex deposition, antiphospholipid antibodies, and infectious factors such as viruses, and it is an important cause of the pathogenesis of SLE [24]. In clinical practice, relative levels of antinuclear antibodies and anti-dsDNA are detected to reflect the disease activity of SLE [26]. Consistently, our findings uncovered that routine blood test data (WBC, HGB, PLT, NE, LY, MPV and RDW) and laboratory indexes (ALB, BUN, Scr, C3, C4, CRP, ESR and anti-dsDNA) were closely linked to the onset of SLE.

The production of proinflammatory factors, cell death and antigen presentation during the progression of SLE are all affected by miRNAs [27-31]. Sheedy et al. [32] discovered 7 differentially expressed miRNAs between SLE patients and healthy controls. Our findings showed that serum level of miR-200a was remarkably downregulated in SLE children. Its level was negatively correlated to SLEDAI, ESR, CRP, BUN and Scr, while it was positively correlated to C3, C4 and ALB in SLE children. In addition, ROC curves demonstrated the diagnostic potential of miR-200a in SLE and LN.

Taken together, this study evaluated the potential interaction between miR-200a and SLE and renal damage. It is concluded that miR-200a level was positively correlated to SLEDAI, which could be an inflammatory indicator for assessing the disease activity of SLE children. Moreover, miR-200a could predict the onset of SLE and the following renal damage.

5. Conclusion

MiR-200a level has a certain correlation to the disease activity of children with initially diagnosed SLE, which can be utilized as an adjuvant indicator in evaluating SLE. Meanwhile, miR-200a has predictive value for SLE-induced renal damage.

Conflict of Interests

The author has no conflicts with any step of the article preparation.

Consent for publications

The author read and approved the final manuscript for publication.

Ethics approval and consent to participate

This study was approved by the ethics committee of Zhanjiang Central People's Hospital.

Informed Consent

Signed written informed consent were obtained from the patients and/or guardians.

Availability of data and material

The data that support the findings of this study are available from the corresponding author upon reasonable request.

Authors' contributions

HZ and MZ designed the study and performed the experiments, XZ collected the data, QL analyzed the data, HZ and MZ prepared the manuscript. All authors read and approved the final manuscript.

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References

1. Khudhair EG, Rebai T, Gazar CAK (2023) Immunomodulatory effects of bone marrow-derived Mesenchymal stem cells on BALB/c mice model with induced Systemic lupus erythematosus (SLE). *Cell Mol Biol* 69:19-24. doi: 10.14715/cmb/2022.69.1.4
2. Tarvin SE, O'Neil KM (2018) Systemic Lupus Erythematosus, Sjogren Syndrome, and Mixed Connective Tissue Disease in Children and Adolescents. *Pediatr Clin North Am* 65:711-737. doi: 10.1016/j.pcl.2018.04.001
3. Hahn BH, McMahon MA, Wilkinson A, Wallace WD, Daikh DI, Fitzgerald JD et al (2012) American College of Rheumatology guidelines for screening, treatment, and management of lupus nephritis. *Arthritis Care Res* 64:797-808. doi: 10.1002/acr.21664
4. Hiraki LT, Benseler SM, Tyrrell PN, Hebert D, Harvey E, Silverman ED (2008) Clinical and laboratory characteristics and long-term outcome of pediatric systemic lupus erythematosus: a longitudinal study. *J Pediatr* 152:550-556. doi: 10.1016/j.jpeds.2007.09.019
5. Yang L, Ma TJ, Zhang YB, Wang H, An RH (2022) Construction and Analysis of lncRNA-miRNA-mRNA ceRNA Network Identify an Eight-Gene Signature as a Potential Prognostic Factor in Kidney Renal Papillary Cell Carcinoma (KIRP). *Altern Ther Health Med* 28:42-51.
6. Li H, Qu L (2022) Correlation of miRNA-21 and blood Cr levels with tumor infiltration and distant metastasis in renal cancer patients. *Cell Mol Biol* 68:117-123. doi: 10.14715/cmb/2022.68.5.16
7. De Guire V, Robitaille R, Tetreault N, Guerin R, Menard C, Bambace N et al (2013) Circulating miRNAs as sensitive and specific biomarkers for the diagnosis and monitoring of human diseases: promises and challenges. *Clin Biochem* 46:846-860. doi: 10.1016/j.clinbiochem.2013.03.015
8. de Planell-Saguer M, Rodicio MC (2013) Detection methods for microRNAs in clinic practice. *Clin Biochem* 46:869-878. doi: 10.1016/j.clinbiochem.2013.02.017
9. Esquela-Kerscher A, Slack FJ (2006) Oncomirs - microRNAs with a role in cancer. *Nat Rev Cancer* 6:259-269. doi: 10.1038/nrc1840
10. Mu YP, Tang S, Sun WJ, Gao WM, Wang M, Su XL (2014) Association of miR-193b down-regulation and miR-196a up-regulation with clinicopathological features and prognosis in gastric cancer. *Asian Pac J Cancer Prev* 15:8893-8900. doi: 10.7314/apjcp.2014.15.20.8893
11. Bruce JP, Liu FF (2014) MicroRNAs in nasopharyngeal carcinoma. *Chin J Cancer* 33:539-544. doi: 10.5732/cjc.014.10175
12. Baumjohann D, Ansel KM (2013) MicroRNA-mediated regulation of T helper cell differentiation and plasticity. *Nat Rev Immunol* 13:666-678. doi: 10.1038/nri3494
13. Singh RP, Massachi I, Manickavel S, Singh S, Rao NP, Hasan S et al (2013) The role of miRNA in inflammation and autoimmunity. *Autoimmun Rev* 12:1160-1165. doi: 10.1016/j.autrev.2013.07.003
14. Miserez AR, Rossi FA, Keller U (1994) Prediction of the therapeutic response to simvastatin by pretreatment lipid concentrations in 2082 subjects. *Eur J Clin Pharmacol* 46:107-114. doi:

- 10.1007/BF00199871
15. Zhao Q, Li M, Chen M, Zhou L, Zhao L, Hu R et al (2016) Lovastatin induces platelet apoptosis. *Environ Toxicol Pharmacol* 42:69-75. doi: 10.1016/j.etap.2016.01.002
 16. Petri M, Orbai AM, Alarcon GS, Gordon C, Merrill JT, Fortin PR et al (2012) Derivation and validation of the Systemic Lupus International Collaborating Clinics classification criteria for systemic lupus erythematosus. *Arthritis Rheum* 64:2677-2686. doi: 10.1002/art.34473
 17. Gladman DD, Ibanez D, Urowitz MB (2002) Systemic lupus erythematosus disease activity index 2000. *J Rheumatol* 29:288-291.
 18. Macedo PA, Garcia CB, Schmitz MK, Jales LH, Pereira RM, Carvalho JF (2012) Juvenile systemic lupus erythematosus and dermatomyositis associated with urticarial vasculitis syndrome: a unique presentation. *Rheumatol Int* 32:3643-3646. doi: 10.1007/s00296-010-1484-4
 19. Kamphuis S, Silverman ED (2010) Prevalence and burden of pediatric-onset systemic lupus erythematosus. *Nat Rev Rheumatol* 6:538-546. doi: 10.1038/nrrheum.2010.121
 20. Malattia C, Martini A (2013) Paediatric-onset systemic lupus erythematosus. *Best Pract Res Clin Rheumatol* 27:351-362. doi: 10.1016/j.berh.2013.07.007
 21. Bennett M, Brunner HI (2013) Biomarkers and updates on pediatric lupus nephritis. *Rheum Dis Clin North Am* 39:833-853. doi: 10.1016/j.rdc.2013.05.001
 22. Birmingham DJ, Irshaid F, Nagaraja HN, Zou X, Tsao BP, Wu H et al (2010) The complex nature of serum C3 and C4 as biomarkers of lupus renal flare. *Lupus* 19:1272-1280. doi: 10.1177/0961203310371154
 23. Kenyon KD, Cole C, Crawford F, Kappler JW, Thurman JM, Bratton DL et al (2011) IgG autoantibodies against deposited C3 inhibit macrophage-mediated apoptotic cell engulfment in systemic autoimmunity. *J Immunol* 187:2101-2111. doi: 10.4049/jimmunol.1003468
 24. Zeng T, Yu J, Tan L, Wu Y, Tian Y, Wu Q et al (2018) Noninvasive indices for monitoring disease course in Chinese patients with autoimmune hepatitis. *Clin Chim Acta* 486:135-141. doi: 10.1016/j.cca.2018.07.030
 25. Tecer D, Sezgin M, Kanik A, Incel NA, Cimen OB, Bicer A et al (2016) Can mean platelet volume and red blood cell distribution width show disease activity in rheumatoid arthritis? *Biomark Med* 10:967-974. doi: 10.2217/bmm-2016-0148
 26. Zivkovic V, Stankovic A, Cvetkovic T, Mitic B, Kostic S, Nedovic J et al (2014) Anti-dsDNA, anti-nucleosome and anti-C1q antibodies as disease activity markers in patients with systemic lupus erythematosus. *Srp Arh Celok Lek* 142:431-436. doi: 10.2217/bmm-2016-0148
 27. Te JL, Dozmorov IM, Guthridge JM, Nguyen KL, Cavett JW, Kelly JA et al (2010) Identification of unique microRNA signature associated with lupus nephritis. *Plos One* 5:e10344. doi: 10.1371/journal.pone.0010344
 28. Tang Y, Luo X, Cui H, Ni X, Yuan M, Guo Y et al (2009) MicroRNA-146A contributes to abnormal activation of the type I interferon pathway in human lupus by targeting the key signaling proteins. *Arthritis Rheum* 60:1065-1075. doi: 10.1002/art.24436
 29. Azizi Dargahlou S, Iriti M, Pouresmaeil M, Goh LPW. (2023) MicroRNAs; their therapeutic and biomarker properties. *Cell Mol Biomed Rep* 3(2): 73-88. doi: 10.55705/cmbr.2022.365396.1085
 30. Kanwal N, Al Samarrai O, Al-Zaidi HMH, Mirzaei A, Heidari M. (2023). Comprehensive analysis of microRNA (miRNA) in cancer cells. *Cell Mol Biomed Rep* 3(2): 89-97. doi: 10.55705/cmbr.2022.364591.1070.
 31. Sasani S, Rashidi Monfared S, Mirzaei AR (2024). Identification of some *Echinophora platyloba* miRNAs using computational methods and the effect of these miRNAs in the expression of TLN2 and ZNF521 genes in different human body organs. *Cell Mol Biomed Rep* 4(1): 43-53. doi: 10.55705/cmbr.2023.386145.1100.
 32. Sheedy FJ, Palsson-McDermott E, Hennessy EJ, Martin C, O'Leary JJ, Ruan Q et al (2010) Negative regulation of TLR4 via targeting of the proinflammatory tumor suppressor PDCD4 by the microRNA miR-21. *Nat Immunol* 11:141-147. doi: 10.1038/ni.1828