

Original Research

Detection of the novel IL-1 family cytokines by QAH-IL1F-1 assay in rheumatoid arthritis

M. Wang^{1,2}, B. Wang², Z. Ma², X. Sun², Y. Tang², X. Li², X. Wu^{3*}

¹ Department of Rheumatology and Immunology, Hebei Medical University Third Affiliated Hospital, 139 Ziqiang Road, Shijiazhuang 050051, Hebei, China

² Department of Immunology, College of Basic Medical Science, Dalian Medical University, Lvshun south Road, Dalian 116044, Liaoning, China

³ Department of Rheumatology and Immunology, Ningbo No.2 Hospital, 41 Xibei Road, Ningbo, 315010, Zhejiang, China

Abstract: The interleukin (IL)-1 family of cytokines comprises 11 members, including 7 pro-inflammatory cytokines (IL-1 α , IL-1 β , IL-18, IL-33, IL-36 α , IL-36 β , IL-36 γ) and 4 anti-inflammatory cytokines (IL-1R antagonist (IL-1Ra), IL-36Ra, IL-37 and IL-38), and play central roles in mediating immune responses. In this study, we detected serum levels of IL-36 subfamily cytokines (including IL-36 α , IL-36 β , IL-36 γ , IL-36Ra and IL-38), IL-37, IL-33 and aimed to investigate the roles of these cytokines in rheumatoid arthritis (RA) preliminarily. A total of 10 RA patients and 10 healthy controls (HCs) were involved in this study, we measured IL-36 subfamily cytokines, IL-37 and IL-33 levels in the serum of the experiment subjects by QAH-IL1F-1 assay. Clinical and laboratory data of the subjects were collected and analyzed by Spearman's rank test. Compared to that of HCs, IL-36 α , IL-36 β , IL-36Ra, IL-38 and IL-33 levels were significantly increased in RA patients. We also found RA patients with elevated IL-36Ra had a higher ESR and RF-IgM, and there was a positive correlation between increased IL-36 α and CRP. Our study suggests that parts of the novel members of IL-1 family cytokines were involved in the pathogenesis of RA, and may provide a novel target for therapies of RA.

Key words: Rheumatoid arthritis, IL-1 family cytokines, QAH-IL1F-1 assay.

Introduction

Rheumatoid arthritis (RA) is a chronic systemic autoimmune disease with unknown aetiology, frequently leading to synovitis and progressive joint damage. However, inflammatory cytokines appear to be involved in the pathogenesis of RA, many of which had not been discovered until now (1). The inhibition of pro-inflammatory cytokines can reduce the manifestations of RA, improve function and retard radiological evidence of joint damage. Cytokine biomarkers may be a useful tool in estimation of the disease activity or joints' damage. Some of the proinflammatory cytokines, including IL-6, TNF- α and IL-17 are currently considered as potential biomarkers (2,3).

The interleukin (IL)-1 family comprises 11 members, namely IL-1 α (IL-1F1), IL-1 β (IL-1F2), IL-1R antagonist (IL-1Ra, IL-1F3), IL-18 (IL-1F4), IL-36R antagonist (IL-36Ra, IL-1F5), IL-36 α (IL-1F6), IL-37 (IL-1F7), IL-36 β (IL-1F8), IL-36 γ (IL-1F9), IL-38 (IL-1F10) and IL-33 (IL-1F11). It is becoming clear that most members of the IL-1 family, such as IL-1 α , IL-1 β and IL-18, play important roles in immune regulation and inflammatory processes by inducing the expressions of other cytokines or matrix metalloproteinases (MMPs) and so on. While some members exert anti-inflammation activities, such as IL-1Ra and IL-36Ra (4,5). The roles of some of IL-1 family members in inflammation and immune modulation were almostly characterized, however, little is to be known about the effects and mechanism of the new members of IL-1 family cytokines, such as IL-36 subfamily cytokines (including IL-36 α , IL-36 β , IL-36 γ , IL-36Ra and IL-38) and IL-37 in RA.

In this study, we detected the levels of these novel

IL-1 family members and aimed to investigate the roles of these cytokines in RA preliminarily. We found IL-36 α , IL-36 β , IL-36Ra, IL-38 and IL-33 levels were significantly increased in RA patients compared to HCs. RA patients with elevated IL-36Ra had a higher erythrocyte sedimentation rate (ESR) and rheumatoid factor (RF)-IgM, and there was a positive correlation between increased IL-36 α and C- reactive protein.

Materials and Methods

Patients and controls

Serum samples were collected from 10 RA patients (Mean \pm SEM age 57.20 \pm 3.72 years) admitted to the ward of the Department of Rheumatology and Immunology, Ningbo No.2 Hospital from June 2014 to August 2014. All RA patients in this study fulfilled the 1987 revised criteria of the American College of Rheumatology (6). Age- and sex-matched healthy controls (HCs, n = 10, Mean \pm SEM age 56.10 \pm 3.94 years) were obtained from the medical examination center. Serum samples were stored at -80°C until used. The study protocol was approved by the ethics committee of Ningbo No.2 Hospital. All study subjects signed written informed consent before participating in the study. All patients had no other autoimmune or systemic diseases.

Received January 20, 2016; Accepted April 11, 2016; Published April 30, 2016

* **Corresponding author:** Xiudi Wu, Department of Rheumatology and Immunology, Ningbo No.2 Hospital, 41 Xibei Road, Ningbo, 315010, Zhejiang, China. Email: wuxiudinbey@163.com

Copyright: © 2016 by the C.M.B. Association. All rights reserved.

Determination of IL-36 subfamily cytokines, IL-37 and IL-33 levels using the QAH-IL1F-1 assay

Serum IL-36 subfamily cytokines, IL-37 and IL-33 levels in all samples were measured with QAH-IL1F-1 assay (Raybiotech, USA) on a GenePix 4000B Microarray Scanner (1311 Orleans Drive Sunnyvale, CA 94089-1136 United States) according to the manufacturer's instructions. The sensitivity was 1 pg/ml.

Clinical and laboratory data analysis

All patients were obtained clinical data on age, sex, disease duration and ESR, CRP, anti-cyclic citrullinated peptides (CCP) antibodies and RF-IgM. ESR was evaluated by the Westergren method. Values ≤ 20.00 mm/h were considered normal. CRP was examined by the immunonephelometry method and a value > 10.00 mg/l was considered positive. Anti-CCP antibody and RF-IgM were tested by enzyme linked immunosorbent assay (ELISA), Anti-CCP antibody with normal ranges of 0-25.00 U/ml, and 0-15.90 IU/ml for RF-IgM.

Statistical analysis

Means \pm standard error of the mean (SEM) were calculated for all conditions, and differences between means were analyzed using Student's t-test and clinical features in RA patients were analyzed by Spearman's rank test. All statistical analyses were performed using GraphPad Prism software (Graph-Pad, San Diego, CA, USA). A P-value < 0.05 was considered significantly different.

Results

Patient clinical characteristics

A total of 10 RA patients were recruited into the study. And they didn't use disease modifying anti-rheumatic drugs(DMARDs). Detailed clinical characteristics and laboratory features of RA were shown in **Table 1**.

Table 1. Clinical and laboratory features in 10 patients with RA.

NO.	Age (yrs)	Sex	Disease duration (yrs)	ESR (mm/h)	CRP (ug/ml)	Anti-CCP (U/ml)	RF-IgM (IU/ml)	Medication within three months
1	76	Female	8	39.00	30.00	783.50	357.00	Meloxicam(15mg/d)
2	72	Female	30	120.00	74.20	1584.00	611.00	Traditional Chinese medicine
3	58	Female	20	30.00	14.20	160.19	11.10	None
4	57	Female	8	98.00	30.00	176.50	399.00	diclofenac slow release tablet(150mg/d)
5	40	Male	5	45.00	45.10	1387.46	621.00	diclofenac slow release tablet(150mg/d)
6	66	Female	1	77.00	30.00	1600.00	90.40	diclofenac slow release tablet(150mg/d) Traditional Chinese medicine
7	53	Female	2	38.00	50.10	45.12	11.10	None
8	41	Female	4	44.00	12.60	11.71	33.60	Ointment for external use
9	53	Female	8	37.00	16.30	456.00	43.30	Traditional Chinese medicine
10	56	Female	2	51.00	8.70	88.70	38.10	Meloxicam(15mg/d)

Table 2. Correlations of the novel IL-1 family cytokines with disease activity and auto-antibody production in RA patients.

	IL-36Ra		IL-36 α		IL-37		IL-36 β		IL-36 γ		IL-38		IL-33	
	r	p	r	p	r	p	r	p	r	p	r	p	r	p
ESR	0.82	0.006**	0.25	0.492	0.44	0.204	-0.26	0.470	0.60	0.073	0.22	0.537	0.25	0.492
CRP	0.10	0.785	0.71	0.027*	0.47	0.166	-0.33	0.349	0.36	0.296	0.58	0.081	0.06	0.892
Anti-CCP	0.28	0.427	0.18	0.632	0.42	0.233	0.22	0.537	0.18	0.632	0.24	0.514	0.14	0.707
RF-IgM	0.74	0.017*	0.47	0.179	0.26	0.470	-0.01	0.973	0.08	0.838	0.41	0.233	0.11	0.759

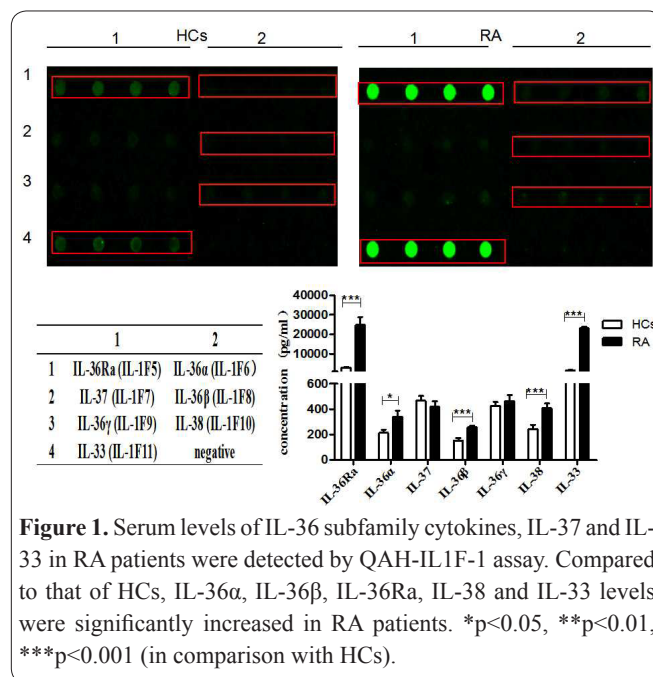


Figure 1. Serum levels of IL-36 subfamily cytokines, IL-37 and IL-33 in RA patients were detected by QAH-IL1F-1 assay. Compared to that of HCs, IL-36 α , IL-36 β , IL-36 γ , IL-38 and IL-33 levels were significantly increased in RA patients. *p<0.05, **p<0.01, ***p<0.001 (in comparison with HCs).

Analysis of serum IL-36 subfamily cytokines, IL-37 and IL-33 levels in RA patients

We measured serum levels of IL-36 subfamily cytokines, IL-37 and IL-33 in RA patients and age-matched HCs by QAH-IL1F-1 assay. The results showed that compared to that of HCs, serum levels of IL-36 α (RA 339.32 \pm 49.07, HCs 214.70 \pm 23.57), IL-36 β (RA 258.20 \pm 13.66, HCs 154.00 \pm 20.62), IL-36 γ (RA 24815.62 \pm 4123.20, HCs 3022.50 \pm 363.63), IL-38 (RA 408.85 \pm 35.82, HCs 234.70 \pm 33.52) and IL-33 (RA 23004.29 \pm 958.11, HCs 1571.00 \pm 132.49) were significantly increased in RA patients. The serum of IL-37 (RA 418.31 \pm 44.88, HCs 466.20 \pm 38.53) and IL-36 γ (RA 463.45 \pm 48.21, HCs 426.10 \pm 30.72) did not differ between RA patients and HCs (**Fig. 1**). Our results suggest that parts of the novel members of IL-1 family cytokines might be associated with the pathogenesis of RA.

Positive correlations of increased IL-36Ra with ESR and RF-IgM and IL-36 α with CRP in RA patients

In our study, the correlations of the increased novel IL-1 family cytokines with clinical activity and auto-antibody levels were analyzed. We found that RA patients with elevated IL-36Ra had higher ESR and RF-IgM, and there was a positive correlation between increased IL-36 α and CRP. But no other correlations were found (Table 2).

Discussion

Cytokines are important mediators in inflammation and immune responses. An imbalance in the cytokine network may lead to inflammatory response and subsequent tissue damage in autoimmune diseases. It is widely recognized that TNF- α , IL-1 β and IL-6 are involved in the pathogenesis of RA and have become main therapeutic targets (7-9). In view of growing need to search for new RA biomarkers, novel cytokine profiles could be used as a powerful predicting tool for indicating the progress of rheumatic disorders (10,11). In this study, we detected serum IL-36 subfamily cytokines, IL-37 and IL-33 levels in RA patients and found that compared to that of HCs, IL-36 α , IL-36 β , IL-36Ra, IL-38 and IL-33 levels were significantly increased in RA patients. Moreover, we found positive correlations of increased IL-36Ra with ESR and RF-IgM, and IL-36 α had a positive correlation with CRP.

IL-1 family of cytokines can be divided into IL-1 subfamily, IL-18 subfamily and IL-36 subfamily. The IL-36 subfamily comprised of IL-36 α , β and γ as well as IL-36 Ra and IL-38 (12). These IL-36 cytokines were mainly expressed in keratinocytes and monocytes / macrophages (13,14). Moreover, IL-36 α and IL-36 β were expressed on T cells (15,16). It has been found that IL-36 can enhance the expression and function of Th17 cytokines in an autocrine manner (17) and promote dendritic cells (DC) to produce inflammatory cytokines at a higher level than other IL-1 family members (18), suggesting its potential role in immunity and inflammatory responses. A recent study found the over-expression of IL-36 α by synoviocytes from patients with RA or psoriatic arthritis (19), Magne (20) showed that joint and serum IL-36 β levels elevated in several samples from RA patients, indicating that IL-36 β can contribute to joint inflammation in RA. Moreover, recent data showed plasma concentrations of IL-36 γ was significantly higher in active systemic lupus erythematosus (SLE) patients than those in HCs (21). In the present study, we found increased serum IL-36 α and IL-36 β levels in RA patients, and there was a positive correlation between increased IL-36 α and CRP, suggesting that IL-36 α / β exert their inflammatory effects in RA.

IL-36Ra acts similar to IL-1Ra, and could exert its antagonistic effects through binding with IL-36R. IL-38 is a recently identified IL-1 family antagonist that functioned similarly to IL-36Ra (22). It was predominantly expressed in the skin and proliferating B-cells of the tonsil (23). Previous studies showed IL-38 gene polymorphisms were associated with psoriatic arthritis (PsA), ankylosing spondylitis (AS) (24-26) and it was showed to be able to suppress IL-17 and IL-22 secretion (22). Interestingly, IL-36Ra also did the same activity

as IL-38, suggesting that IL-38 and IL-36Ra are partial receptor antagonists. Consistent with previous reports, in our results, there were higher serum levels of IL-36Ra and IL-38 in RA patients. And we found positive correlation of increased IL-36Ra with ESR and RF-IgM in RA patients. As IL-1 family members, higher IL-38 and IL-36Ra suggest they might be a negative feedback increase to inhibit inflammatory responses in RA.

IL-37 was expressed in monocytes, macrophages and epithelial cells (27). Zhao *et al.* reported that plasma IL-37 level was significantly higher in RA patients, and positively correlated with IL-17A, TNF- α and disease activity (28). But in our experiment, perhaps because of limited samples, no significant difference of serum IL-37 levels was found between RA patients and HCs. More patients need be detected to testify the exact role of IL-37 in RA.

IL-33 was widely expressed in different cell types, such as macrophages, DC, fibroblasts, endothelial cells. IL-33 binding with its ligand ST2L on mast cells, macrophages and activated neutrophils, enhanced joint inflammation by inducing cytokines such as TNF- α , IL-1 β , IL-6, IL-17, and interferon (IFN)- γ in arthritis models (29,30). Yeon-Sik Hong (31) found that serum level of IL-33 correlated with that of IL-1 β and IL-6. Serum IL-33 and sST2 levels decreased together with CRP, after DMARDs therapy in patients with treatment-naive RA. Another study observed that IL-33 induced neutrophil migration by activating synoviocytes and macrophages and could directly attract neutrophils to the site of inflammation (32). Here we also confirmed that IL-33 were significantly increased in RA patients which might explain the pro-inflammatory role of IL-33 in rheumatic disorders.

In this study, we preliminary revealed the association between the new members of IL-1 family cytokines and RA patients. However, our study had several limitations. First, the sample size was not large. Another limitation was the heterogeneity of our patients. Therefore, more patients need to be examined to confirm the roles of these cytokines in RA patients. Further studies of the new member of IL-1 family cytokines would be interesting to help with understanding the pathogenesis of RA.

Acknowledgments

This work was supported by the National Natural Science Foundation of China (No. 81172847, 81373214); Liaoning Province Natural Science Foundation (2014022013); Nanjing Young Medical Talent Project; China Postdoctoral Science Foundation the First Class (2012M510073) and the Natural Science Foundation of Ningbo (No.2013A610259).

References

1. Feldmann M and Maini SR Role of cytokines in rheumatoid arthritis: an education in pathophysiology and therapeutics. *Immunological Reviews*, 2008, 223(1): 7-19.
2. Miao J, Zhang K, Lv M, Li Q, Zheng Z, Han Q, Guo N, Fan C, Zhu P. Circulating Th17 and Th1 cells expressing CD161 are associated with disease activity in rheumatoid arthritis. *Scand J Rheumatol*, 2014, 43(3): 194-201.
3. Niu X and Chen G. Clinical biomarkers and pathogenic-related cytokines in rheumatoid arthritis. *J Immunol Res* 2014, 698192.

4. Dinarello CA. Immunological and inflammatory functions of the interleukin-1 family. *Annu. Rev. Immunol.*, 2009, 27:519-550.
5. Dinarello CA. The IL-1 family and inflammatory diseases. *Clin Exp Rheumatol*, 2002, 20:S1-13.
6. Arnett FC, Edworthy SM, Bloch DA, Mcshane DJ, Fries JF, Cooper NS, Healey LA, Kaplan SR, Liang MH, Luthra HS, Medsger TA, Mitchell DM, Neustadt DH, Pinals RS, Schaller JG, Sharp JT, Wilder RL and Hunder GG. The American Rheumatism Association 1987 revised criteria for the classification of rheumatoid arthritis. *Arthritis Rheum.*, 1998, 31:315-324.
7. Olsen NJ and Stein CM. New drugs for rheumatoid arthritis. *The New England Journal of Medicine*, 2004, 350(21):2167-2226.
8. Bansard C, Lequerre T, Derambure C, Vittecoq Q, Hiron M, Daragon A, Pouplin S, Daveau M, Boyer Q, Tron F, Le Loet X and Salier JP. Gene profiling predicts rheumatoid arthritis responsiveness to IL-1Ra (anakinra). *Rheumatology*, 2011, 50(2):283-292.
9. Smolen JS, Aletaha D, Koeller M, Weisman MH and Emery P. New therapies for treatment of rheumatoid arthritis. *The Lancet*, 2007, 370(9602):1861-1874.
10. Meyer PW, Hodkinson B, Ally M, Musenge E, Wadde AA, Fickl H, Tikly M and Anderson R. Circulating cytokine profiles and their relationships with autoantibodies, acute phase reactants, and disease activity in patients with rheumatoid arthritis. *Mediators Inflamm*, 2010, 2010:158514.
11. Deane KD, O'Donnell CI, Hueber W, Majka DS, Lazar AA, Derber LA, Gilliland WR, Edison JD, Norris JM, Robinson WH and Hokers VM. The number of elevated cytokines and chemokines in pre-clinical seropositive rheumatoid arthritis predicts time to diagnosis in an age-dependent manner. *Arthritis Rheum.*, 2010, 62(11):3161-3172.
12. Sims JE and Smith DE. The IL-1 family: regulators of immunity. *Nat Rev Immunol*, 2010, 10:89-102.
13. Smith DE, Renshaw BR, Ketchum RR, Kubin M, Garka KE and Sims JE. Four new members expand the interleukin-1 superfamily. *J. Biol. Chem.*, 2000, 275:1169-1175.
14. Barksby HE, Nile CJ, Jaedicke KM, Taylor JJ and Preshaw PM. Differential expression of immunoregulatory genes in monocytes in response to *Porphyromonas gingivalis* and *Escherichia coli* lipopolysaccharide. *Clin. Exp. Immunol.*, 2009, 156:479-487.
15. Vigne S, Palmer G, Lamacchia C, Martin P, Talabot-Ayer D, Rodriguez E, Ronchi F, Sallusto F, Dinh H, Sims JE and Gabay C. IL-36R ligands are potent regulators of dendritic and T cells. *Blood*, 2011, 118: 5813-5823.
16. Li Y, Messina C, Bendaoud M, Fine DH, Schreiner H, Tsiagbe VK. Adaptive immune response in osteoclastic bone resorption induced by orally administered *Aggregatibacter actinomycetemcomitans* in a rat model of periodontal disease. *Mol. Oral Microbiol.*, 2010, 25: 275-292.
17. Carrier Y, Ma H.L., Ramon H.E., Napierata L, Small C, O'Toole M, Young D.A., Fouser L.A., Nickerson-Nutter C, Collins M, Dunussi-Joannopoulos K, Medley QG. Inter-regulation of Th17 cytokines and the IL-36 cytokines in vitro and in vivo: Implications in psoriasis pathogenesis. *J. Investig. Dermatol.*, 2011, 131, 2428-2437.
18. Vigne S, Palmer G, Lamacchia C, Martin P, Talabot-Ayer D, Rodriguez E, Ronchi F, Sallusto F, Dinh H, Sims J.E, Gabay C. IL-36R ligands are potent regulators of dendritic and T cells. *Blood*, 2011, 118, 5813-5823.
19. Frey S, Derer A, Messbacher ME, Baeten DL, Bugatti S, Montecucco C, Schett G and Hueber AJ. The novel cytokine interleukin-36 α is expressed in psoriatic and rheumatoid arthritis synovium. *Ann Rheum Dis*, 2013, 72:1569-1574.
20. Magne D, Palmer G, Barton JL, Mezin F, Talabot-Ayer D, Bas S, Duffy T, Noger M, Gueme PA, Nicklin MJ and Gabay C. The new IL-1 family member IL-1F8 stimulates production of inflammatory mediators by synovial fibroblasts and articular chondrocytes. *Arthritis Res Ther*, 2006, 8(3):R80.
21. Chu M, Wong CK, Cai Z, Dong J, Jiao D, Kam NW, Lam CW and Tam LS. Elevated Expression and Pro-Inflammatory Activity of IL-36 in Patients with Systemic Lupus Erythematosus. *Molecules*, 2015, 20:19588-19604.
22. van de Veerdonk FL, Stoeckman AK, Wu G, Boeckermann AN, Azam T, Netea MG, Joosten LA, van der Meer JW, Hao R, Kalabokis V and Dinarello CA. IL-38 binds to the IL-36 receptor and has biological effects on immune cells similar to IL-36 receptor antagonist. *Proceedings of the National Academy of Sciences of the United States of America*, 2012, 109(8): 3001-3005.
23. Lin H, Ho AS, Haley-Vicente D, Zhang J, Bernal-Fussell J, Pace AM, Hansen D, Schweighofer K, Mize NK and Ford JE. Cloning and characterization of IL-1HY2, a novel interleukin-1 family member. *J. Biol. Chem.*, 2001, 276:20597-20602.
24. Rahman P, Sun S, Peddle L, Snelgrove T, Melay W, Greenwood C and Gladman D. Association between the interleukin-1 family gene cluster and psoriatic arthritis. *Arthritis and Rheumatism*, 2006, 54(7):2321-2325.
25. Chou CT, Timms AE, Wei JC, Tsai WC, Wordsworth BP and Brown MA. Replication of association of IL1 gene complex members with ankylosing spondylitis in Taiwanese Chinese. *Annals of the Rheumatic Diseases*, 2006, 65(8):1106-1109.
26. Guo ZS, Li C, Lin ZM, Huang JX, Wei QJ, Wang XW, Xie YY, Liao ZT, Chao SY and Gu JR. Association of IL-1 gene complex members with ankylosing spondylitis in Chinese Han population. *International Journal of Immunogenetics*, 2010, 37(1):33-37.
27. Buefler P, Azam T, Gamboni-Robertson F, Reznikov LL, Kumar S, Dinarello CA and Kim SH. A complex of the IL-1 homologue IL-1F7b and IL-18-binding protein reduces IL-18 activity. *Proc Natl Acad Sci USA*, 2002, 99:13723-13728.
28. Zhao PW, Jiang WG, Wang L, Jiang ZY, Shan YX and Jiang YF. Plasma levels of IL-37 and correlation with TNF- α , IL-17A, and disease activity during DMARD treatment of rheumatoid arthritis. *PloS One*, 2014, 9:e95346.
29. Xu D, Jiang HR, Kewin P, Li Y, Mu R, Fraser AR, Pitman N, Kurowska-Stolarska M, Mckenzie AN, McInnes IB and Liew FY. IL-33 exacerbates antigen-induced arthritis by activating mast cells. *Proc Natl Acad Sci USA*, 2008, 105:10913-10918.
30. Palmer G, Talabot-Ayer D, Lamacchia C, Toy D, Seemayer CA, Viatte S, Finckh A, Smith DE and Gabay C. Inhibition of interleukin-33 signaling attenuates the severity of experimental arthritis. *Arthritis Rheum*, 2009, 60:738-749.
31. Yeon-Sik Hong, Su-Jin Moon, Young-Bin Joo, Chan-Hong Jeon, Mi-La Cho, Ji Hyeon Ju, Hye-Jwa Oh, Yu-Jung Heo, Sung-Hwan Park, Ho-Youn Kim and Jun-Ki Min. Measurement of Interleukin-33 (IL-33) and IL-33 Receptors (sST2 and ST2L) in Patients with Rheumatoid Arthritis. *Immunology, Allergic Disorders & Rheumatology*, 2011, 26: 1132-1139.
32. Waldiceu A Verri Jr, Fabrício O Souto, Silvio M Vieira, Sergio C L Almeida, Sandra Y Fukada, Damo Xu, Jose C Alves-Filho, Thiago M Cunha, Ana T G Guerrero, Rafaela B Mattos-Guimaraes, Fabiola R Oliveira, Mauro M Teixeira, João S Silva, Iain B McInnes, Sergio H Ferreira, Paulo Louzada-Junior, Foo Y Liew, Fernando Q Cunha. IL-33 induces neutrophil migration in rheumatoid arthritis and is a target of anti-TNF therapy. *Ann Rheum Dis*, 2010, 69:1697-1703.